

## RESEARCH ARTICLE SUMMARY

## PLATELETS

# Platelets sequester extracellular DNA, capturing tumor-derived and free fetal DNA

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Full article and list of author affiliations:  
<https://doi.org/10.1126/science.adp3971>

**INTRODUCTION:** Platelets are the smallest and second most abundant cell type in our blood, contributing to a range of physiological processes, including hemostasis, vascular maintenance, and innate immunity. Whereas megakaryocytes, the platelet-producing cells in the bone marrow, have a large, highly polyploid nucleus, platelets have no nucleus at all and have traditionally been assumed to be devoid of DNA. Platelets have a distinct cellular physiology with a series of surface-connected membrane channels, called the open canalicular system, mediating efficient internalization and release of biomolecules, including nucleic acids such as viral RNA and DNA, within the circulation.

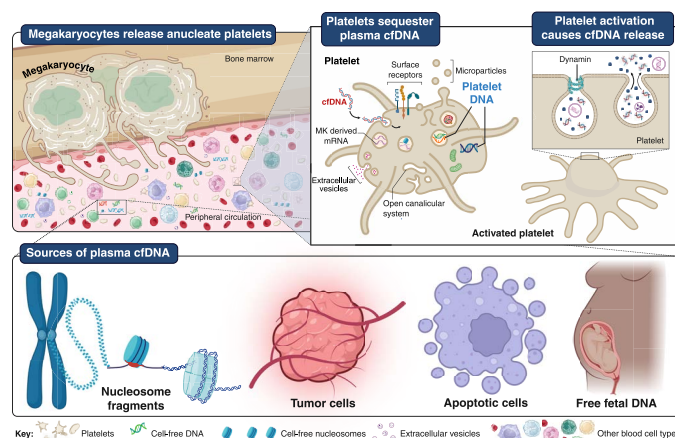
**RATIONALE:** Given their known ability to sense and internalize exogenous nucleic acids, we hypothesized that platelets may sequester endogenous extracellular DNA during circulation. Conceivably, this process might contribute to physiological mechanisms that limit the abundance of cell-free DNA (cfDNA) in plasma, which is immunostimulatory. This phenomenon would be relevant to liquid biopsy approaches, many of which focus on cfDNA isolated from platelet-depleted plasma, in which platelet nucleic acids are discarded. cfDNA contained within platelets may offer an untapped, additional reservoir of clinically relevant genetic material to, for example, increase sensitivity in cancer screening to detect low-tumor burden disease.

**RESULTS:** By using high resolution imaging and flow cytometry, we confirmed that platelets contain fractional quantities of DNA. To test whether platelets can acquire DNA after their release from megakaryocytes, we performed live cell imaging and coculture experiments, demonstrating transfer of mutant DNA from malignant cells to healthy donor platelets *in vitro*. We confirmed that cfDNA sequestration by platelets occurs *in vivo*, corroborating both visually (with fluorescence *in situ* hybridization) and molecularly (with droplet digital polymerase chain reaction) that platelets from healthy pregnant females carrying male babies contained fragments of the Y chromosome.

Whole-genome sequencing of platelet DNA revealed a fragmentation pattern and nucleosome footprint that mirrored the attributes of cfDNA isolated from platelet-depleted plasma. The DNA fragments in platelets were histone-associated chromatin, with DNA methylation marks suggesting a similar cellular composition to cfDNA with contributions from blood leukocytes and megakaryocytes and a minor contribution from hepatocytes in healthy individuals.

Given our hypothesis that platelets clear cfDNA from plasma, we reasoned that plasma cfDNA would increase in the setting of thrombocytopenia. Indeed, acute platelet depletion in an immune thrombocytopenic purpura (ITP) mouse model led to a rise in plasma cfDNA, and patients with ITP and severely low platelet counts exhibited significantly higher plasma cfDNA levels than individuals with normal platelet counts.

We found that platelets could acquire DNA by internalizing both DNA-loaded extracellular vesicles released by apoptotic cells and



**Platelets sequester cfDNA during circulation.** Bone marrow megakaryocytes release platelets into the bloodstream. During circulation, platelets encounter extracellular DNA—histone-bound nucleosome fragments—released by cells undergoing physiological apoptosis, including tumor cells and free fetal DNA. Platelets protect internalized DNA from external DNase degradation and release their DNA cargo in a dynamin-dependent manner upon platelet activation. [Figure created with BioRender.com]

membrane-free DNA fragments. Uptake and release of DNA could be pharmacologically manipulated. Platelet-encapsulated DNA was protected from external deoxyribonuclease (DNase) degradation, and inhibiting platelet exocytosis increased the abundance of DNA obtained from platelets.

Lastly, we demonstrated that tumor-derived DNA was present in platelets of patients with advanced cancer, where cfDNA was in high abundance; in low-tumor burden disease; and also in those with premalignant colonic polyps.

**CONCLUSION:** Platelets can acquire a repertoire of DNA fragments that map across the nuclear genome, including DNA derived from tumor cells and free fetal DNA. Prior work has shown that tumor cell-derived RNA is detectable within circulating platelets. Given their abundance, ease of isolation, and tissue-wide perfusion, platelets are ideally positioned to serve as biosensors for genetic perturbations across tissues. This finding has broad clinical relevance and sets the stage for future work to clarify the role of platelets in cfDNA homeostasis as well as the fate and physiological consequences of DNA fragments released at sites of platelet activation. □

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## PLATELETS

# Platelets sequester extracellular DNA, capturing tumor-derived and free fetal DNA

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Platelets are anucleate blood cells vital for hemostasis and immunity. During cell death and aberrant mitosis, nucleated cells release DNA, resulting in “cell-free” DNA in plasma (cfDNA). An excess of cfDNA is deleterious. Given their ability to internalize pathogen-derived nucleic acids, we hypothesized that platelets may also clear endogenous cfDNA. We found that, despite lacking a nucleus, platelets contained a repertoire of DNA fragments mapping across the nuclear genome. We detected fetal DNA in maternal platelets and cancer-derived DNA in platelets from patients with premalignant and cancerous lesions. As current liquid biopsy approaches utilize platelet-depleted plasma, important genetic information contained within platelets is being missed. This study establishes a physiological role for platelets that has not previously been highlighted, with broad translational relevance.

Platelets are fundamental to hemostasis, vascular maintenance, and both innate and adaptive immunity. Platelets inherit RNAs from their parent megakaryocytes together with translational and splicing machinery. As part of their role in immunity, platelets sense pathogen-derived nucleic acids and internalize DNA and RNA viruses, triggering inflammatory responses (1–4). Notably, in addition to pathogen-derived nucleic acids, RNAs originating from solid tumor cells and inhaled pine tree pollen have also been detected in human platelets (5, 6), indicating that platelets internalize RNA from a range of cellular sources during peripheral circulation. Although platelet RNA has emerging utility as a liquid biopsy approach for hematological (7) and

solid malignancies (8, 9), whether platelets contain any form of DNA—and if so, its genomic origin—has not been extensively investigated.

During cell death and aberrant mitosis, nucleated cells release chromosomal DNA, resulting in “cell-free” DNA in plasma (cfDNA). Analysis of cfDNA from blood plasma is currently implemented in a wide range of clinical settings, including cancer care (10–16) and prenatal testing (17). A major obstacle to clinical utility is the low sensitivity in scenarios where the abundance of target DNA is low. We hypothesized that in addition to internalizing pathogen-derived exogenous DNA, platelets may clear endogenous cfDNA from plasma, including DNA fragments released by healthy cells as well as those bearing oncogenic mutations. As conventional methods isolate cfDNA from platelet-depleted plasma, any DNA fragments that are sequestered within platelets would be missed. We therefore also hypothesized that including DNA captured by platelets may add value in diagnostic pathways.

## Results

### Platelets contain DNA

To determine whether DNA is present within human platelets, platelets were purified from three sources: whole peripheral blood, isolated plasma, and “plateletpheresis” concentrates, clinical-grade platelet pools isolated from healthy donors for transfusion purposes (Fig. 1A). We examined platelets by fluorescence microscopy after labeling purified platelets with NUCLEAR-ID Red DNA, a highly specific, cell-permeable dye that intercalates with double-stranded DNA (dsDNA). A proportion of platelets showed a clear signal for the DNA probe (Fig. 1B). To verify that this represented nuclear DNA, we confirmed the presence of a nuclear genomic region of the *TP53* gene in platelet DNA (pDNA) (fig. S1, A and B). Immunofluorescence staining of a peripheral blood smear also showed a clear signal for the DNA stain Hoechst in platelets, confirming that the DNA was not acquired by platelets during purification from whole blood (fig. S1C).

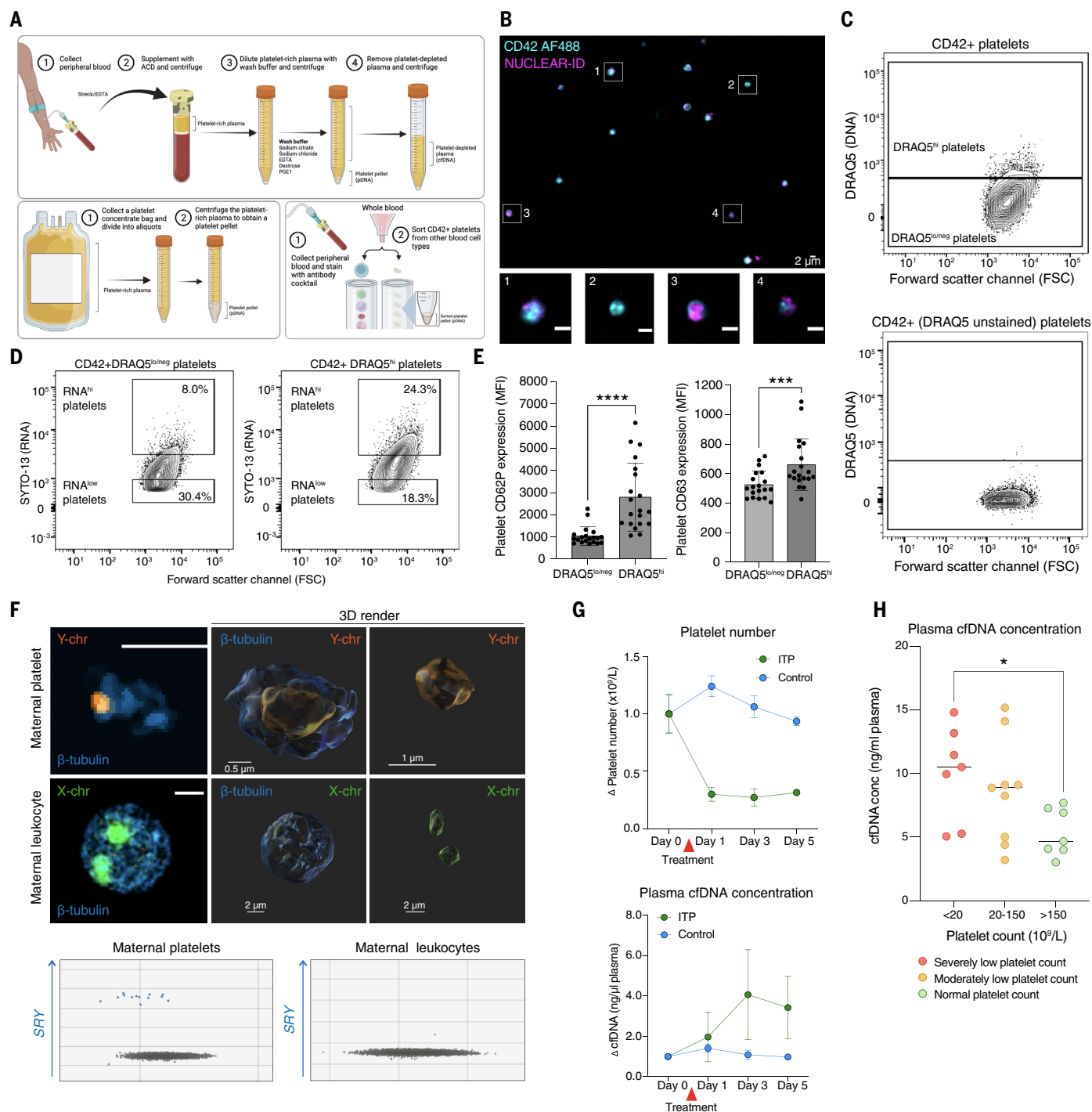
To quantify the proportion of platelets that contained DNA we used flow cytometry, examining positivity for DRAQ5, a dsDNA-specific dye. Most platelets showed low-level DRAQ5 positivity, whereas around 8% were strongly positive (DRAQ5<sup>hi</sup>) (Fig. 1C and fig. S1D). Prior studies have shown that newly produced or “younger” platelets can be distinguished from “older” platelets by their higher content of RNA and that younger platelets also have higher expression of markers of platelet receptor activation or degranulation (18–20). Costaining for RNA and DNA showed that the majority of DRAQ5<sup>hi</sup> platelets were low or negative for RNA (Fig. 1D and fig. S1E), indicating that DNA is present in both newly produced (RNA-high) and older (RNA-low) platelets. Comparing DRAQ5<sup>hi</sup> and DRAQ5<sup>lo</sup> platelets showed that DRAQ5<sup>hi</sup> platelets contained more RNA and had higher expression of markers of activation than DRAQ5<sup>lo/neg</sup> platelets, implying that the DNA content of platelets also decreased during platelet lifespan and after platelet activation, in parallel with RNA content (Fig. 1, D and E).

To investigate platelet content of DNA in a larger cohort, we extracted pDNA from peripheral blood of 76 adult donors (tables S1 to S4). The extremely high purity of the platelet pellets was confirmed by flow cytometry analysis and cytospin preparations (fig. S1, A, F, and G). DNA was obtained from both the platelet pellet and platelet-depleted plasma (cfDNA) in all cases, with a higher yield of DNA from platelet pellets than from matched platelet-depleted plasma (i.e., a conventional cfDNA preparation). In this cohort, a median abundance of 7.18 genomic copies was isolated from platelets per microliter of blood plasma, which equates to approximately 0.000012 genomic equivalents of DNA per platelet (fig. S1H).

### Platelets sequester DNA during circulation

To determine whether the DNA detected in platelets was captured during peripheral circulation or solely derived from parent megakaryocytes (21), we collected antenatal blood samples and looked for fragments

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**Fig. 1. Platelets sequester DNA during circulation.** (A) Schematic showing sources of purified platelets. (Top) Sequential centrifugation of peripheral blood to simultaneously extract pDNA and cDNA. (Bottom left) Centrifugation of healthy donor plateletpheresis concentrates to obtain a platelet pellet. (Bottom right) FACS of whole blood. (B) Healthy donor-derived platelets identified by CD42 AF488 (cyan) show an internal fluorescent signal for NUCLEAR-ID, a cell-permeable dye that intercalates with dsDNA (magenta). White boxes mark magnified regions; representative images are shown below. Scale bars, 2  $\mu$ m. (C) Flow cytometric analysis of CD42<sup>+</sup> healthy donor-derived platelets stained with and without the DNA marker DRAQ5. (Top) Eight percent of circulating platelets stain strongly positive for DRAQ5;  $n = 20$  healthy donors concatenated into a single FCS file. (Bottom) Single CD42<sup>+</sup>-stained platelets show no DRAQ5 autofluorescence;  $n = 10$  healthy donors; concatenated data shown. (D) Flow cytometric analysis quantifying RNA content (SYTO-13 positivity) of CD42<sup>+</sup>, DRAQ5<sup>hi</sup>, and DRAQ5<sup>lo/neg</sup> platelets ( $n = 10$  healthy donors; concatenated data shown). (E) Platelets with a higher DNA content (DRAQ5<sup>hi</sup>) showed higher levels of activation as determined by P-selectin (CD62P) and CD63 expression. Each dot represents an individual healthy donor.  $n = 19$  and 20 donors for CD63 and CD62P, respectively. Bars show mean  $\pm$  SD. \*\*\* $P < 0.001$ ; \*\*\*\* $P < 0.0001$ ; as determined by a Wilcoxon signed-rank test. (F) FISH and ddPCR show detection of the Y-chromosome gene SRY in maternal platelets but not in leukocytes sampled from mothers of male offspring prior to delivery. Platelets and leukocytes were counterstained with  $\beta$ -tubulin (blue) and imaged using a ZEISS LSM900, 63 $\times$  magnification. Representative images and ddPCR data are shown; 3D rendering was performed using ImarisViewer.  $n = 5$  and 25 female donors for FISH and ddPCR, respectively. (G) Rise in cfDNA following acute depletion of platelets in healthy mice following administration of an antiplatelet antibody.  $n = 5$  mice per day;  $n = 20$  mice per condition. Each dot shows

mean  $\pm$  SEM fold change at days 1, 3, and 5 compared with baseline (day 0, untreated mice). (H) Plasma cfDNA concentrations (nanograms per milliliter) for patients with ITP and severely ( $<20 \times 10^9/L$ ,  $n = 7$ ) or moderately ( $20 \times 10^9$  to  $150 \times 10^9/L$ ,  $n = 9$ ) low platelet counts or counts in the normal range ( $\geq 150 \times 10^9/L$ ,  $n = 7$ ). \* $P < 0.05$ ; for a Mann-Whitney  $U$  test, each point shows data from one individual, with line at the median. MFI, mean fluorescence intensity.

of the Y chromosome in platelets of mothers with uncomplicated pregnancies carrying male offspring (table S5). Using Y chromosome fluorescence in situ hybridization, Y chromosome fragments were visualized in the platelets of mothers carrying a male fetus (Fig. 1F and movie S1). In 100% of samples ( $n = 30$ ), we correctly ascertained the sex of the fetus by the presence or absence of the Y chromosome gene sex-determining region Y (*SRY*) alleles in maternal platelets (Fig. 1F). *SRY* remained detectable in platelets at 24 and 48 hours after delivery of the baby (fig. S1I) and was not detected in mononuclear cells (MNCs) (Fig. 1F) or red blood cells (RBCs) (fig. S1J) from the same blood samples, indicating that free fetal DNA could be detected exclusively in maternal platelets. This suggested that the detection of *SRY* in maternal platelets was not due to “contaminating” cfDNA in the cell pellets nor the presence of fetal blood cells in the maternal circulation. Although cross-placental transfer of fetal platelets cannot be fully excluded, this observation confirmed the presence of nuclear DNA within platelets.

### Platelets “clear” cfDNA from plasma

We reasoned that if platelets were clearing cfDNA from plasma, then the abundance of cell- or platelet-free DNA in plasma would increase if the platelet count was reduced. To test this, we induced acute and specific antibody-mediated depletion of platelets in mice. In accordance with our hypothesis, induction of immune thrombocytopenia led to an acute rise in plasma cfDNA concentration (Fig. 1G). Similarly, we found higher concentrations of plasma cfDNA in patients with immune thrombocytopenic purpura (ITP) who had severely low platelet counts ( $<20 \times 10^9/L$ ) as compared to those with normal platelet counts (Fig. 1H and table S6;  $P < 0.05$ ,  $n = 23$ ).

Together, these data indicate a biological role for platelets in the clearance of cfDNA from plasma.

### Platelets rapidly capture DNA from nucleated cells

To confirm that platelets sequestered chromosomal fragments released from nucleated cells, we used live cell imaging to visualize DNA uptake in vitro. Colorectal adenocarcinoma cells (COLO205) were labeled with DNA probes and washed twice to remove any noninternalized probe prior to incubation with platelets. Rapid acquisition of COLO205 cell-derived DNA by platelets was observed (Fig. 2, A and B; fig. S2, A to C; and movies S2 and S3). The average DNA fluorescence intensity of the 168 platelets imaged plateaued at  $\sim 6$  min, suggestive of saturation of DNA uptake (Fig. 2C).

### DNA is internalized and released on platelet activation

To determine the localization of pDNA, we performed high-resolution Airyscan imaging of healthy donor platelets spread on collagen-coated glass coverslips. During spreading, platelets evaginate the membranes of their open canalicular system (OCS), an internal surface-connected system of channels, to increase their surface area (22). This process extrudes materials captured within the OCS and pushes intracellular contents to the center (22). Imaging of spread platelets costained for P-selectin (CD62P), a marker of platelet activation and degranulation, and DRAQ5 revealed that DNA was located at the center of spread platelets (Fig. 2D). Calculation of the DRAQ5 staining intensity during progressive computational erosion of the CD62P mask from the edge of the platelet to the center further confirmed that DNA was internalized within the body of the platelet and not held within the OCS (fig. S2D).

We next performed colocalization analyses by costaining platelets for DNA (DRAQ5), the different types of platelet storage granules (alpha and dense granules), and other organelle markers. We observed

strong colocalization between DNA and membrane-bound internal vesicles (CD81 and LAMP-2), with no correlation with mitochondria (TOM20) (Fig. 2, E to H), confirming that the DNA content of platelets was largely contained within membrane-bound vesicles and not stored in granules or restricted to mitochondrial DNA.

### Platelets internalize DNA-loaded extracellular vesicles

DNA-loaded extracellular vesicles (EVs) are released during cellular apoptosis, and viable cells also remove excess cytoplasmic DNA created during aberrant cycles of mitosis by the release of DNA-loaded EVs (23–27). Platelets internalize EVs through their OCS and endocytosis (28), and platelet uptake of tumor cell-derived EVs has previously been shown to transfer cancer cell-derived protein cargo and mRNA transcripts (29).

We therefore investigated whether EVs may mediate DNA transfer to platelets. Platelets isolated from female donors were incubated with EVs derived from male Burkitt's lymphoma (BL2) cells or isogenic BL2 cells engineered to overexpress the apoptosis-suppressing gene *BCL2* (BL2-Bcl-2 cells). This system enabled us to also explore whether DNA loading of EVs occurred during apoptosis. EVs were isolated from BL2 and BL2-Bcl-2 cells after ultraviolet irradiation, generating apoptotic EVs (apo EVs) from BL2 cells or nonapoptotic (nonapo EVs) from BL2-Bcl-2 cells. The EVs were treated with deoxyribonuclease (DNase) to remove any noninternalized DNA (Fig. 2I). DNA-loaded EVs were visible adherent to, as well as within, platelets following incubation (Fig. 2, J and K; movies S4 and S5; and fig. S2, E and F). Copies of *SRY* were detectable in platelets from female donors only following incubation with BL2-derived EVs as well as in higher abundance following incubation with apo EVs than with nonapo EVs (Fig. 2L and fig. S2G). Similarly, by using whole chromosome X and Y fluorescence in situ hybridization (FISH) probes, fragments of both X and Y chromosomes were visualized in female platelets following incubation with HEL cells that also derive from a male donor (Fig. 2M).

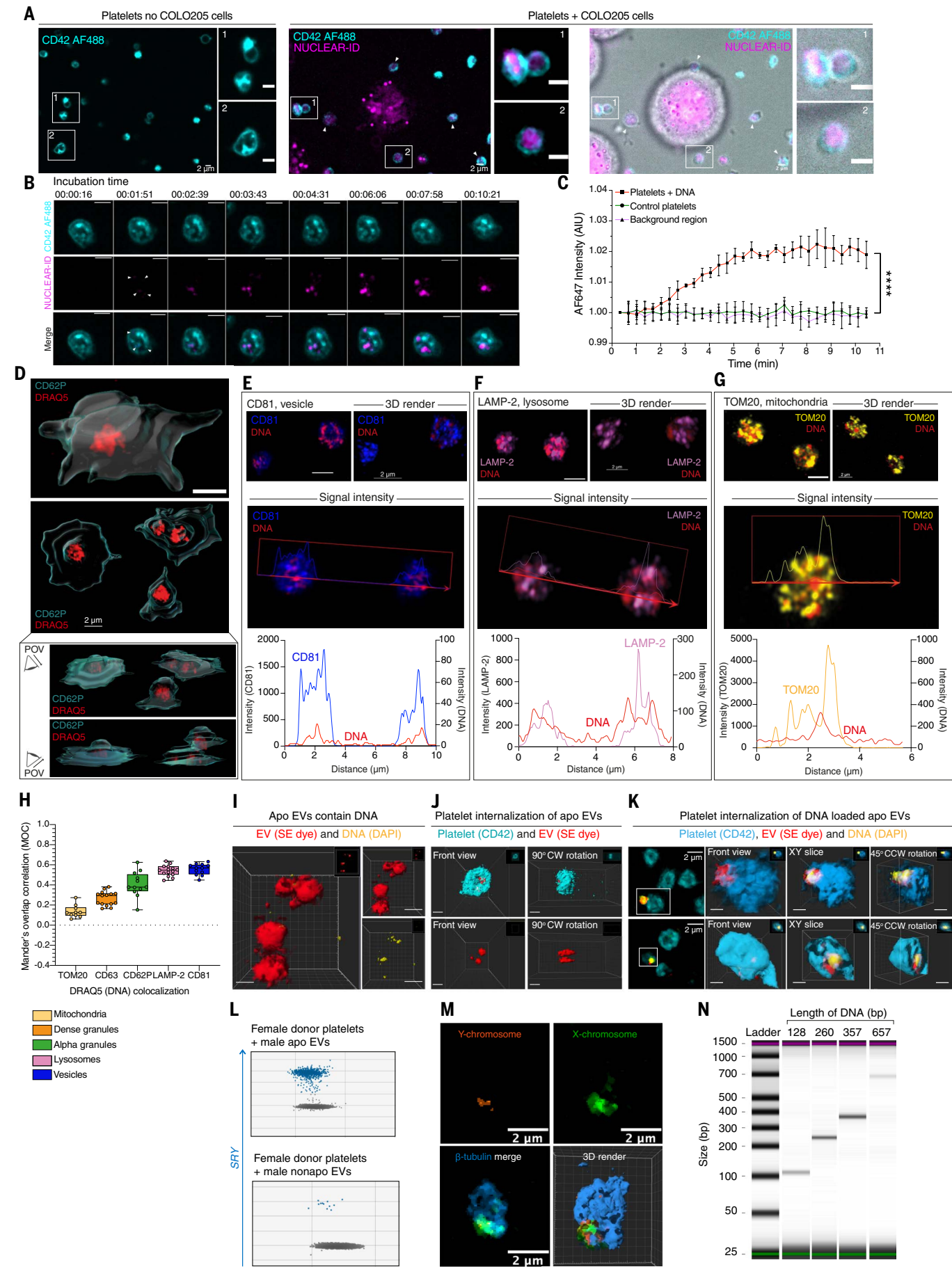
### Platelets internalize membrane-free DNA fragments

To test whether platelets could also internalize “free” DNA fragments that were not contained within EVs, we exposed healthy donor platelets to synthetic DNA fragments ranging from 128 to 657 base pairs (bp) in length. Platelets efficiently internalized all tested DNA fragments, confirming that platelets sequestered non-membrane-bound DNA of varying lengths in addition to DNA-loaded EVs (Fig. 2N).

### Pharmacological modulation of DNA trafficking

To explore whether DNA uptake and release by platelets could be pharmacologically manipulated, we tested the impact of agonists and inhibitors known to block platelet activation and/or specific components of cargo trafficking. Activation of platelets with the canonical platelet agonist thrombin receptor activating peptide (TRAP) decreased their DNA content (Fig. 3A), indicating that DNA is released by platelets upon activation. In keeping with this, the yield of DNA isolated from platelets was higher when platelets were treated with two potent inhibitors of platelet activation: aspirin and ibrutinib, a Bruton's tyrosine kinase (BTK) inhibitor (Fig. 3, B and C, and fig. S3A).

We next tested a range of inhibitors known to block specific components of cargo trafficking by platelets, including inhibitors of actin (cytochalasin D), membrane ruffling (imipramine), clathrin (Pitstop 2), caveolae-dependent endocytosis (nystatin), dynamin (Dynole) and  $\text{Na}^+/\text{H}^+$  exchange or micropinocytosis (amiloride). In keeping with previous literature suggesting that platelets “engulf” EVs through endocytosis



**Fig. 2. Platelets rapidly internalize DNA released by nucleated cells through uptake of DNA-loaded EVs and non-membrane-encapsulated DNA fragments.**

(A) Healthy donor-derived, CD42 AF488 (cyan)-labeled platelets before (left) and after (middle and right) coincubation with COLO205 cells labeled with NUCLEAR-ID (magenta) to track DNA transfer. White boxes show magnified regions; arrows highlight platelet uptake of COLO205-derived DNA. Scale bars, 2  $\mu$ m. Middle (fluorescence) and far right (bright field) are the same image. (B) Live-cell time-lapse imaging of a representative platelet showing internalization of DNA. Scale bars, 3  $\mu$ m. (C) Time-course quantification of pDNA fluorescence intensity; data points show mean  $\pm$  SD. AF647 signal was measured in background regions to detect fluctuations. \*\*\*\* $P$  < 0.0001, as determined by a Wilcoxon signed-rank test. Tracking shown for platelets incubated with control media ( $n$  = 168 platelets) or media conditioned by NUCLEAR-ID-labeled COLO205 cells ( $n$  = 173 platelets). (D to G) Representative images acquired using the Airyscan detector of a ZEISS LSM980 as well as 3D rendering performed using ImarisViewer. (D) Healthy donor platelets spread on collagen and stained with CD62P, to create a translucent cell surface mask (turquoise), and DRAQ5 (DNA, red). Four representative platelets are shown. (Bottom) Alternate views of the same platelets from above and below; scale bars, 2  $\mu$ m. [(E) to (G)] (Top) Representative images and 3D renders of two healthy donor platelets showing colocalization of pDNA (DRAQ5, red) with (E) CD81 (vesicles, blue), (F) LAMP-2 (lysosomes, pink), and (G) TOM20 (mitochondria, yellow). (Bottom) Pixel intensity ( $y$  axis) along defined lines (distance in micrometers;  $x$  axis) was analyzed using ZEISS Zen software. (H) Manders' correlation coefficients were calculated for DNA with mitochondria (TOM20), dense granules (CD63), alpha granules (CD62P), lysosomes (LAMP-2), and vesicles (CD81). Box plots show the median, first quartile, and third quartile, with whiskers extending to minimum and maximum values. Platelets analyzed per marker: TOM20 ( $n$  = 10), CD63 ( $n$  = 15), CD62P ( $n$  = 12), LAMP-2 ( $n$  = 15), and CD81 ( $n$  = 12). (I) A representative 3D reconstruction of EV clusters released by apoptotic BL2 cells labeled with an amine-reactive membrane dye (CF658, red) and DNA stain (DAPI, yellow), revealing internalized DNA. Imaged using a ZEISS LSM900, 63 $\times$  magnification. Scale bars, 2  $\mu$ m. (J) Platelets stained with CD42 AF488 (cyan) following incubation with labeled EVs (red) showing three EVs within a single representative platelet. 3D modeling shows 90° clockwise rotation. Scale bar, 1.4  $\mu$ m. (K) 3D reconstruction showing platelet internalization of DNA-loaded apo EVs (DNA, yellow; EVs, red). Two representative platelets are shown: one with surface-bound DNA-loaded EVs (top) and one with internalized EVs (bottom). Scale bars, 0.5  $\mu$ m unless stated otherwise. (L) ddPCR quantification of the Y-chromosome gene *SRY* in DNA derived from platelets of a female donor after incubation with apo EVs and nonapo EVs. Representative data from  $n$  = 3 independent experiments. (M) Representative FISH and 3D render demonstrating fragments of X and Y chromosomes in female donor platelets following incubation with male HEL cells. Platelets were counterstained with  $\beta$ -tubulin (blue). Imaged using a ZEISS LSM900, 63 $\times$  magnification; 3D render was created using ImarisViewer. (N) Electrophoresis data showing recovery of 128- to 657-bp synthetic DNA fragments from washed healthy donor platelets after overnight incubation. Representative data from  $n$  = 5 independent experiments. AIU, arbitrary intensity units; BL2, Burkitt's lymphoma cells; chr, chromosome; CCW, counterclockwise; CW, clockwise; POV, perspective of view; SE, succinimidyl ester.

at the OCS membrane, we found that platelet uptake of DNA-loaded apo EVs was reduced by the dynamin inhibitor Dynole, indicating that DNA-loaded apo EVs were internalized by clathrin-mediated endocytosis (Fig. 3, E and F).

These inhibitors did not reduce the uptake of free DNA fragments by platelets, suggesting that this occurs through an alternative, clathrin-independent mechanism (Fig. 3G). Rather, we found that Dynole and amiloride substantially increased the amount of DNA obtained from the platelets exposed to synthetic DNA fragments (Fig. 3, H and I). Dynole and amiloride also block platelet exocytosis and the release of intracellular contents (30, 31). Accordingly, we found that Dynole reduced pDNA release (Fig. 3, J to L).

Together, these data indicated platelet activation was not required for DNA uptake, but that DNA was released by platelets upon activation. Furthermore, platelet content of DNA can be pharmacologically modulated to increase the abundance of DNA preserved within platelets (fig. S3B).

### Detecting cancer mutations in platelets

Given the growing importance of cfDNA detection in cancer screening and molecular profiling, we sought to determine whether platelets may take up tumor-derived DNA fragments that conferred cancer-specific genetic variants.

Platelets from healthy donors were incubated with a variety of human cancer cell lines harboring common cancer-associated gene mutations, including colorectal cells and erythroleukemia cells (Fig. 4A). Analysis of DNA isolated from platelets following coincubation with cancer cell lines detected all relevant oncogenic mutations, including *KRAS*<sup>G12D</sup>, *BRAF*<sup>V600E</sup>, *PIK3CA*<sup>H1047R</sup>, and *JAK2*<sup>V617F</sup> (Fig. 4, B and C). No cancer-specific gene mutations were identified in healthy donor platelets incubated alone, but wild-type alleles were detected (Fig. 4C), indicative of DNA sequestered by platelets prior to blood sampling.

To determine whether the ability to capture extracellular DNA was specific to platelets or shared by other blood cell types, we tested whether *BRAF*<sup>V600E</sup> derived from COLO205 cells was also sequestered in vitro by peripheral blood MNCs and RBCs. Although mutant *BRAF*<sup>V600E</sup> was readily detectable in pDNA (Fig. 4D), fewer copies of *BRAF*<sup>V600E</sup> were detected in MNCs and RBCs (Fig. 4D and fig. S3, C to E), where they were of negligible abundance in comparison to wild-type alleles (fig. S3, D and E).

### pDNA is protected from nucleases

To test whether pDNA was protected from external nuclease digestion, conditioned media (analogous to cfDNA) and platelets were treated with DNase following conditioning by HEL cells that carry the *JAK2*<sup>V617F</sup> mutation. Detection of *JAK2*<sup>V617F</sup> in HEL cell-conditioned media was completely abrogated by DNase treatment (Fig. 4E). By contrast, there was no reduction in the abundance of *JAK2*<sup>V617F</sup> in platelets following DNase treatment (Fig. 4E), indicating that the majority of DNA was encapsulated by the platelet outer membrane and protected from enzymatic degradation. This was verified by using synthetic DNA fragments and target-specific primers (fig. S3F).

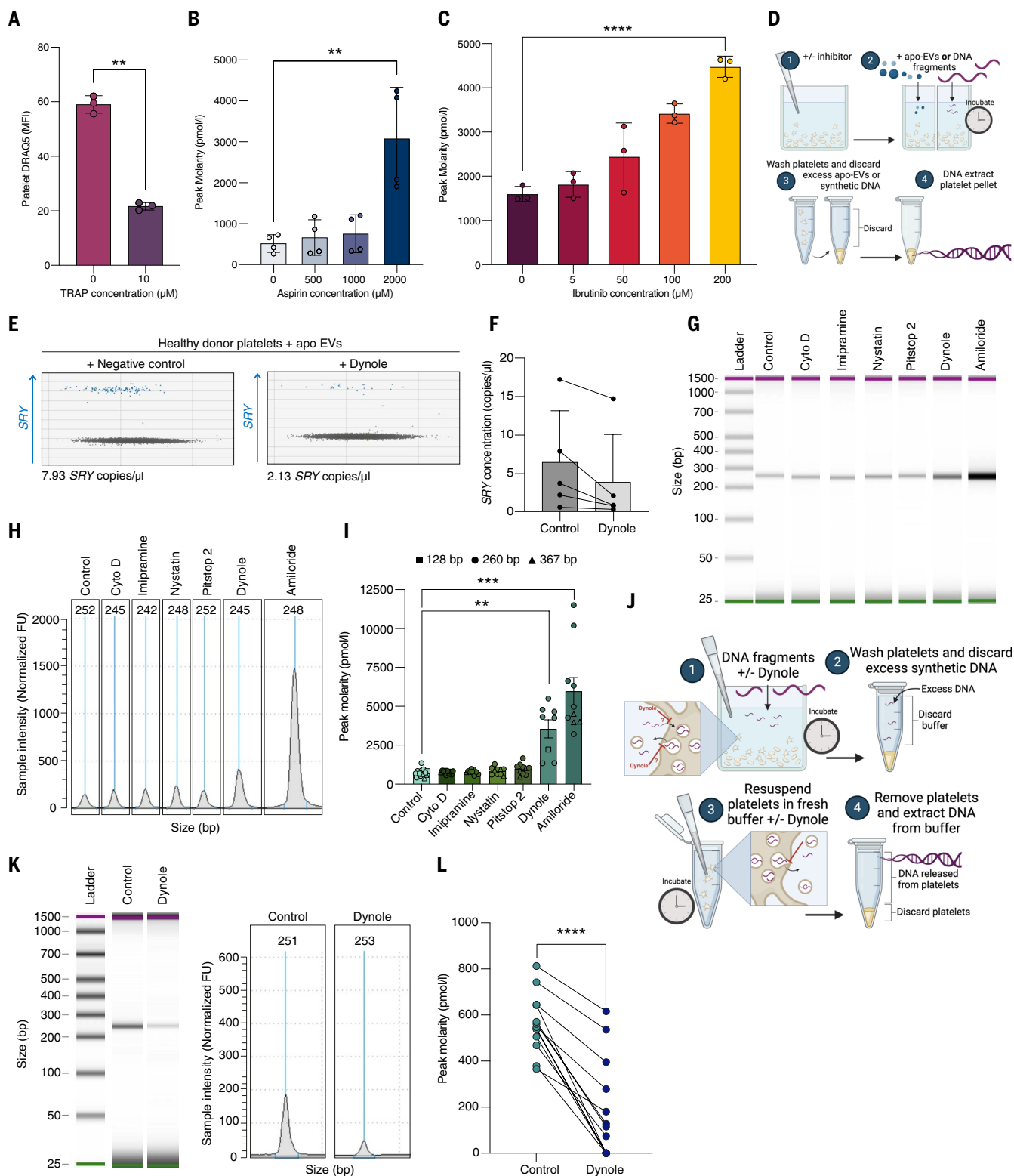
### Genomic distribution and origin of pDNA

We then set out to explore the genomic characteristics of pDNA and validate its utility for detection of cancer gene aberrations in vivo.

cfDNA has a classical fragmentation profile with a periodicity reflecting nucleosomal footprints (12). Electrophoretic analysis of pDNA showed fragments of around 12,000 to 16,000 bp, and many samples also showed shorter fragments of around 120 to 500 bp with nucleosome footprints, similar to the nucleosome footprint of cfDNA (fig. S4, A and B). Low-input chromatin immunoprecipitation (ChIP) for H3k27 histone acetylation of DNA isolated from platelet pellets and live-cell imaging of platelets coincubated with H2B-mCherry-labeled HEL cells confirmed that the DNA sequestered by platelets was associated with nucleosomes and that histone modifications are preserved in pDNA (fig. S4, C and D, and movie S6).

To compare the genomic origin of pDNA and cfDNA, whole-genome sequencing (WGS) was performed on matched pDNA and cfDNA extracted from the peripheral blood of 15 donors—of whom 10 had active malignancy, and 5 had undergone surgery and/or chemoradiotherapy—to screen for residual disease (table S4). Size selection was performed on the pDNA to separate the shorter (<600 bp) “s-pDNA,” which were similar in length to cfDNA fragments, from the longer fragments (“l-pDNA”).

We performed low-pass WGS (mean target coverage of 1X) on s-pDNA, l-pDNA, and paired cfDNA ( $n$  = 15). Platelets contain a small number of mitochondria, estimated at four to six mitochondria per cell (32, 33). The majority of fragments from both s- and l-pDNA mapped to the nuclear genome (97.0 to 99.9% for s-pDNA and 69.9 to 99.1% for l-pDNA) (Fig. 4F). As expected, given that the mitochondrial



**Fig. 3. pDNA content can be pharmacologically modulated.** (A) Flow cytometric analysis of DNA (DRAQ5) content in healthy donor-derived platelets, with and without activation using 10  $\mu$ M TRAP-6.  $n = 3$ ; points represent data from individual healthy donors; bars show mean  $\pm$  SD; \*\* $P < 0.01$ , as determined by a paired  $t$  test. (B and C) Inhibition of platelet activation with (B) aspirin or (C) ibrutinib increases the yield of DNA from platelet pellets.  $n = 3$  to 4 independent experiments; plots show mean  $\pm$  SD peak molarity corresponding to DNA TapeStation peaks for synthetic DNA fragments coincubated with platelets prior to washing and DNA extraction. \*\* $P < 0.01$ ; \*\*\*\* $P < 0.0001$ , as determined by an unpaired  $t$  test. (D) Schematic of the method for investigating DNA uptake. Platelets were pretreated with inhibitors, incubated with DNA-loaded apo EVs or synthetic DNA fragments, then washed, pelleted, and subjected to DNA extraction. (E and F) ddPCR quantification of SRY in platelet-derived DNA from female donors after incubation of platelets with apo EVs treated with or without Dynole. Fewer SRY copies were detected in dynamin-inhibited platelets compared with controls. (E) Representative ddPCR plot and (F)  $n = 5$  independent experiments, paired line plot shows mean  $\pm$  SD, with lines connecting data points before and after

inhibition. **(G and H)** Representative electrophoresis showing recovery of 260-bp synthetic DNA fragments from platelets treated with inhibitors known to block components of cargo trafficking. More DNA was extracted from platelets treated with Dynole and amiloride. **(I)** Summary plot showing mean  $\pm$  SD for all DNA fragment lengths (128 to 367 bp) following treatment with inhibitors [(G) and (H)]. Data from four independent experiments ( $n = 4$ ) with two to five replicate wells per condition.  $**P < 0.01$ ;  $***P < 0.001$ ; as determined by a paired  $t$  test. **(J)** Schematic of the method for investigating pDNA release. Platelets were treated with or without Dynole, incubated with synthetic DNA, washed, resuspended in fresh buffer, and incubated for 3 hours. DNA was then extracted from platelet-depleted buffer. **(K)** Representative electrophoresis and **(L)** a paired-line plot showing significant inhibition of pDNA release following Dynole treatment, with lines connecting data points before and after inhibition. Data from five independent experiments ( $n = 5$ ), with two to three replicate wells per condition.  $****P < 0.0001$ ; as determined by a paired  $t$  test. cyto D, cytochalasin D; FU, fluorescence units.

genome is  $\sim 16,500$  bp, a higher subfraction (0.9 to 30.1%) of l-pDNA mapped to the mitochondrial genome versus 0.081 to 2.968% of the shorter fragments (Fig. 4F).

Mapping across the genome revealed that pDNA fragments covered the entire nuclear genome (fig. S4E), mirroring cfDNA and demonstrating that sampling of pDNA would enable genetic variants across the genome to be detected. Compared with cfDNA, the s-pDNA fragments showed more even coverage, as indicated by a lower Gini index (34) (fig. S4F).

### pDNA maps to the nuclear genome, and coverage mirrors cfDNA

We then obtained deeper sequencing (mean sequencing depth  $15\times \pm 3$  versus  $12\times \pm 1$  for cfDNA and s-pDNA, respectively) of paired cfDNA and s-pDNA, including five from donors after therapy (samples 1 to 5) and two from donors with active, untreated pancreatic and colorectal carcinoma (table S4). No notable differences were detected in sequencing quality (base quality, 3' bias, mapping rate) between s-pDNA and cfDNA, except for a higher estimated polymerase chain reaction (PCR) duplication rate (cfDNA,  $3.2\% \pm 0.6\%$ ; pDNA,  $11.3\% \pm 7.2\%$ ), indicating lower complexity of short pDNA libraries, presumably due to the size selection.

Nucleosome-bound cfDNA is more protected from degradation than nucleosome-free DNA. This leads to a characteristic fragment-length distribution of cfDNA, with mono- and dinucleosome length peaks at approximately 167 and 320 bp, respectively (35). The peak mean fragment length for s-pDNA was  $166 \text{ bp} \pm 0.8$ , with a second, dinucleosome peak at approximately 328 bp. The dinucleosome peak was, by a fraction, larger in s-pDNA than cfDNA (fig. S5A), likely reflecting the increased protection of pDNA from nuclease digestion, as observed in our *in vitro* experiments (Fig. 4E and fig. S3F).

cfDNA fragments of tumor cell origin are shorter than wild-type fragments (36, 37). In accordance with this, we observed that both mono- and dinucleosome fragment lengths were notably shorter in both pDNA and cfDNA in sample 6, derived from a patient with an inoperable pancreatic tumor and containing readily detectable tumor-derived cfDNA, indicating that fragmentomic analysis may be applied to pDNA to detect the presence of cancer (38) and that platelets capture DNA fragments that resemble those found in cfDNA *in vivo* (Fig. 4G and table S4).

To confirm the capture of circulating tumor cell-derived DNA (ctDNA) by platelets *in vivo*, we used ichorCNA (39) to look for copy number alterations (CNAs) and estimate the fraction of tumor-derived DNA. Clonal amplifications and deletions in chromosomes 6 and 7 as well as subclonal amplifications in chromosome 2 were detected in both cfDNA and s-pDNA from sample 6 (from the patient with pancreatic cancer), with an estimated tumor fraction of 25% (Fig. 4H). CNAs detected in s-pDNA mirrored those present in cfDNA, and the estimated proportion of tumor-derived DNA was highly correlated between s-pDNA and cfDNA (fig. S5, B and C). Selecting DNA fragments smaller than 150 bp increased the estimated tumor content of s-pDNA from 25 to 30%, in parallel to the increase seen in cfDNA (fig. S5D). No CNAs were detected in the l-pDNA from sample 6 (fig. S5E), in line with previous reports that ctDNA is enriched in shorter fragments (37), nor in either the cfDNA or pDNA from patients after therapy.

Matched tumor and healthy tissue biopsy samples were available for sample 7 from a patient with colorectal carcinoma, enabling us to look deeper for low-level ctDNA by screening for tumor-specific mutations not present in matched healthy tissue. We compared the detection rate of tumor-specific mutations from the relevant donor with detection of random samples of mutations taken from other patients with colorectal carcinoma (40). The matched tumor mutations were detected at a higher rate than the random mutations in the cfDNA and s-pDNA and at similar frequency (likelihood rate of  $8.92 \times 10^{-11}$  for cfDNA and  $2.30 \times 10^{-7}$  for s-pDNA), with lower-level detection in the l-pDNA ( $P < 0.0004$ ; Fig. 4I), confirming the presence of ctDNA in platelets.

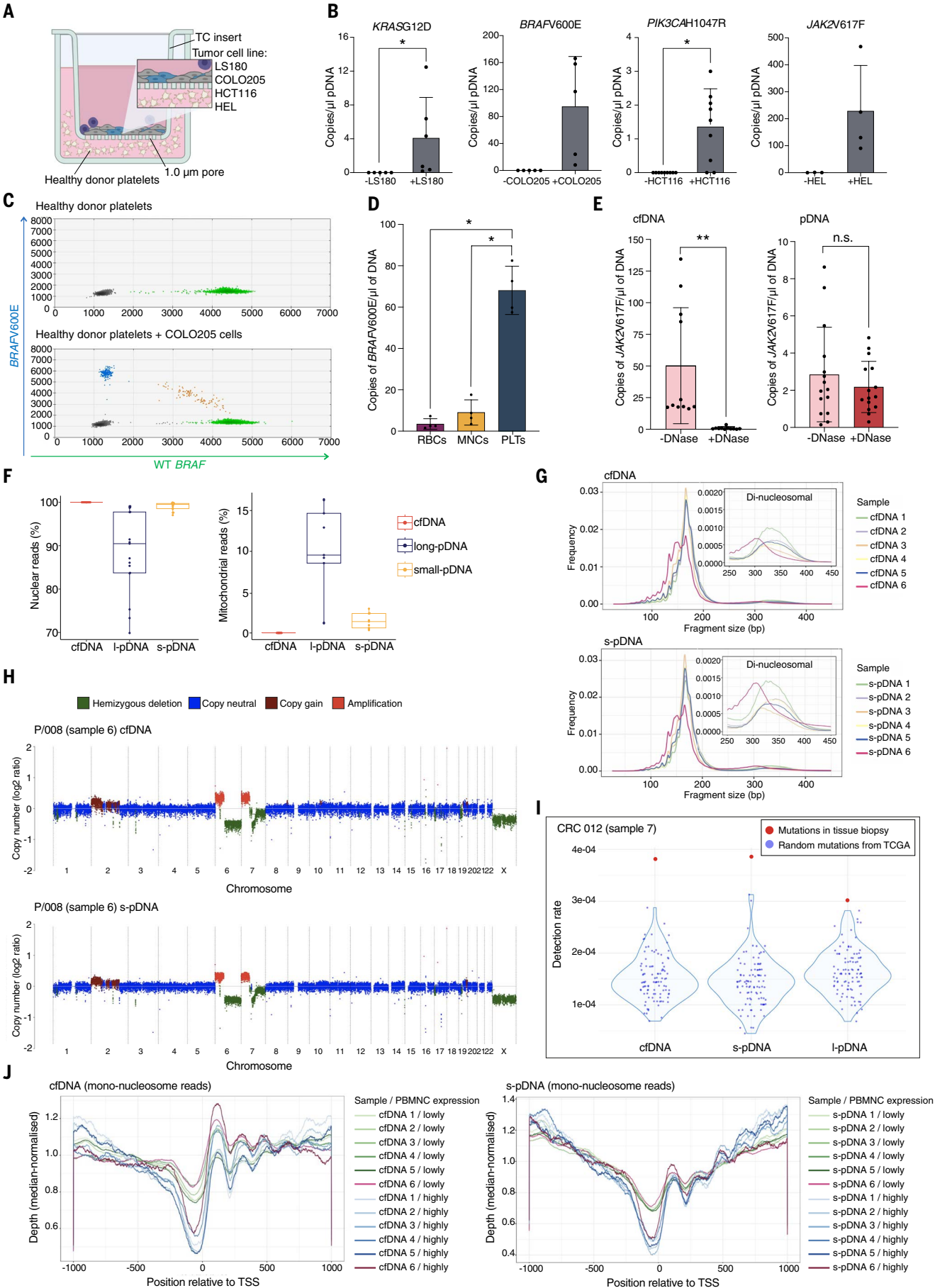
These data indicated that ctDNA was present within platelets in similar abundance to cfDNA, both in patients with high levels of ctDNA and clear CNAs ( $>20\%$ , sample 6; Fig. 4H) and in those with extremely low levels ( $<0.5\%$ , sample 7; Fig. 4I), where targeted analysis and matched tumor tissue was required. Overall, ctDNA was identified in both platelets and cfDNA in 20% of the pretreatment samples either by detecting CNAs or known tumor mutations, in keeping with the sensitivity of WGS for ctDNA detection reported in large clinical cohorts (41).

The enrichment for nucleosome-bound DNA in cfDNA has previously been used to gain insight into gene expression patterns and cell-of-origin, with prior reports showing that a depletion of reads occurs over transcriptional start sites (TSSs) with periodicity in coverage relative to the TSS (42). In addition, DNA methylation studies have shown that the majority ( $>90\%$ ) of cfDNA in both healthy individuals and in those with cancer is of blood cell origin, including neutrophils, monocytes, erythroblasts (43), and megakaryocytes (44). We found very similar depletion in sequencing coverage and periodicity around TSS for genes that are highly expressed in peripheral blood MNCs in both pDNA and cfDNA, suggesting a shared cellular origin of pDNA and cfDNA (Fig. 4J and fig. S5F). Notably, the depletion in coverage for high-expression MNC genes was less marked for the sample that had an estimated ctDNA fraction of 25% than for the other samples (Fig. 4J).

To orthogonally validate the cell-of-origin of pDNA, whole-genome methylation analysis was performed using the bisulfite-free sequencing method, TET-assisted pyridine borane sequencing (TAPS) (45). The data was then deconvoluted using a published atlas of cell-specific methylation patterns (46). This confirmed a similar cellular origin of pDNA and cfDNA, with the majority originating from blood leukocytes and a minor contribution from hepatocytes. Of note, a higher proportion of l-pDNA than s-pDNA mapped to megakaryocyte-specific loci (35.9 versus 16.8%), suggesting that more l-pDNA may derive from parent megakaryocytes (fig. S5G).

### High abundance of mutant alleles in platelets in colorectal and pancreatic carcinoma

To test whether including an analysis of pDNA would add value to plasma cfDNA in the setting of cancer diagnostics, we used a mouse model of colorectal carcinoma in which expression of *KRAS*<sup>G12D</sup> and *TP53* mutations are induced by the villin promoter, resulting in locally invasive colorectal adenocarcinoma with a low incidence of metastases



**Fig. 4. Platelets contain a repertoire of DNA fragments that map over the human nuclear genome, including tumor-derived DNA-bearing cancer-associated gene mutations.** (A) Schematic of the experimental system for coincubating healthy donor platelets with malignant cells, separated by 1  $\mu\text{m}$  membrane inserts to allow extracellular biomolecule and small EV exchange while preventing cell transfer. After coincubation, platelets were removed, washed three times, and subjected to DNA extraction. (B) Number of copies of mutant alleles detected per microliter of DNA extracted from platelets incubated with (+) or without (–) colorectal (LS180, COLO205, and HCT116) and erythroleukemia (HEL) cell lines. Data were collected from one to three independent experiments, with three to five replicate wells per condition. Each point represents one coculture experiment per condition. Replicates per condition were as follows: LS180 ( $n = 2$  experiments, 3 replicates each), COLO205 ( $n = 1$  experiment, 4 and 5 replicates), HCT116 ( $n = 3$  experiments, 3 replicates each), and HEL ( $n = 1$  experiment; 3 and 4 replicates). Bars represent mean  $\pm$  SD;  $*P < 0.05$ , as determined by a Wilcoxon paired signed-rank test. (C) Representative ddPCR analysis showing quantification of wild-type (WT) *BRAF* and *BRAF*<sup>V600E</sup> alleles in pDNA extracted from healthy donor platelets before (top) and after coincubation with COLO205 colorectal cancer cells (bottom). (D) Quantification of *BRAF*<sup>V600E</sup> in RBCs, MNCs, and platelets (PLTs) following incubation in media conditioned by COLO205 cells. Data are presented as mean  $\pm$  SD; individual points represent  $n = 2$  independent experiments, each with 2 replicates.  $*P < 0.05$ ; as determined by a Mann-Whitney *U* test. (E) Impact of DNase treatment on the detection of *JAK2* mutant alleles in pDNA and cfDNA (media) following coincubation of healthy donor platelets with HEL cells. Data are presented as mean  $\pm$  SD; individual points represent  $n = 5$  independent experiments, each with 3 replicates. n.s., not significant;  $**P < 0.01$ ; determined by a Wilcoxon signed-rank test. (F) Percentage of fragments mapping to the nuclear and mitochondrial genomes from cfDNA, short ( $>100$  and  $<600$  bp) and long ( $>600$  bp) pDNA fragments. In the box plot, the center line shows the median, whiskers indicate the minimum and maximum values, and each data point represents one patient;  $n = 15$  donors with gastrointestinal carcinoma. (G) Fragment length distribution of paired, aligned reads for cfDNA (top) and s-pDNA (bottom).  $n = 6$  donors with gastrointestinal carcinoma; samples 1 to 5 show a primary peak at  $\sim 165$  bp (mononucleosomal) and a smaller secondary peak at  $\sim 325$  bp (dinucleosomal; inset). Sample 6 (red line), from a patient with untreated pancreatic adenocarcinoma, has shorter mono- and dinucleosomal fragments than samples 1 to 5 (post-anticancer therapy patients). (H) Deviation from median coverage in 100-kbp windows across all chromosomes for cfDNA (top) and s-pDNA (bottom) for P/008 (sample 6), revealing chromosomal aberrations in chromosomes 2, 6, and 7 (copy number gains and amplifications shown in red; deletions shown in green). (I) Detection rates of mutations in cfDNA, s-pDNA, and l-pDNA for sample 7 where matched tumor biopsy was available. The red dot indicates mutations detected in the tumor biopsy; blue dots indicate 100 random samples of mutations from other patients with colorectal carcinoma from The Cancer Genome Atlas (TCGA) dataset. (J) Read-depth distribution around transcription start sites (TSSs) of genes highly [transcripts per million (TPM)  $>15$ , blue] and lowly (TPM  $<15$ , green) expressed in peripheral blood MNCs for cfDNA (left) and s-pDNA (right) mononucleosome reads.  $n = 6$  donors with gastrointestinal carcinoma. Depth per position per sample was normalized to the median read depth across all genes in that expression category. Single-letter abbreviations for the amino acid residues referenced throughout the paper and figures are as follows: G, Gly; D, Asp; V, Val; E, Glu; H, His; R, Arg; F, Phe.

and mice with *KRAS*<sup>G12D/+</sup>, *TP53*<sup>fl/fl</sup>, plus *NOTCH1*<sup>fl/+</sup>, with more aggressive and metastatic disease (47) (Fig. 5A). pDNA and cfDNA were extracted simultaneously from the same blood sample (Fig. 5A).

Mutant *KRAS*<sup>G12D</sup> as well as wild-type *KRAS* alleles were readily detectable in both cfDNA and platelets of mice with colorectal carcinoma. In more than half of the mice (13/20, 65%), copies of *KRAS*<sup>G12D</sup> per microliter of DNA were in higher abundance in pDNA than cfDNA, both in mice with locally invasive and metastatic disease (Fig. 5, B to E).

To corroborate these findings in a second model and examine an early stage during tumorigenesis prior to tumor dissemination, we used an orthotopic pancreatic cancer model, where pancreatic cancer cells were inserted within a Matrigel plug into the body of the pancreas (48). Fourteen days after implantation and prior to local tumor invasion, *KRAS*<sup>G12D</sup> alleles were readily detected in murine platelets (Fig. 5C and fig. S6, A and B) and were not reduced in mice receiving low-dose systemic aspirin therapy (fig. S6C).

### *BRAF*<sup>V600E</sup> in platelets of patients with precancer

Improving the sensitivity for liquid biopsy screening is crucial in the setting of low-tumor burden disease and in patients with premalignant lesions, where the abundance of ctDNA is low. Sessile serrated lesions (SSLs) are high-risk, premalignant colon polyps driven by mutations in *BRAF* that account for around one-quarter of colorectal cancers (CRCs) (49). To explore the utility of pDNA for detecting premalignant lesions, we collected platelets and cfDNA from patients found to have SSLs at colonoscopy ( $n = 31$ ) versus controls ( $n = 14$ ).

Mutant *BRAF*<sup>V600E</sup> alleles were detected in either the cfDNA and/or pDNA in 16.1% of the patients with serrated polyps (5/31) (Fig. 5F), consistent with previous reports of the frequency of detectable ctDNA in SSL cohorts (50, 51). Mirroring our findings in the CRC mouse model, around half (3/5) of the patients with detectable ctDNA had a higher number of *BRAF*<sup>V600E</sup> copies in pDNA than in cfDNA (Fig. 5, F to H, and fig. S6D). *BRAF*<sup>V600E</sup> was not detected in any of the pDNA samples from 14 controls (Fig. 5F), demonstrating high specificity of targeted mutational analysis of pDNA in this setting and that at least half of the relevant ctDNA is sequestered by blood platelets.

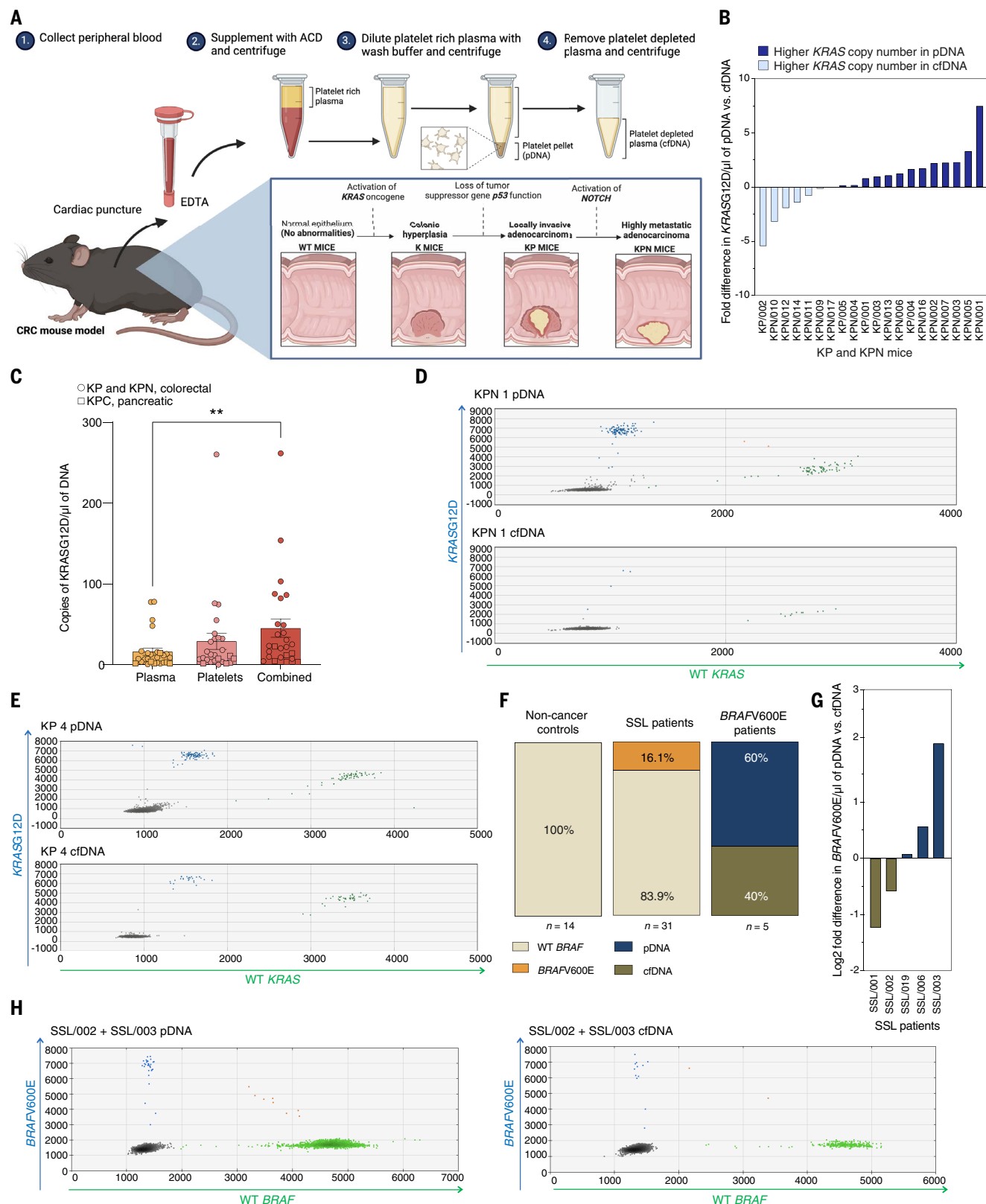
## Discussion

Prior studies of platelet nucleic acid content have only focused on RNA (8, 52–54), although the presence of DNA in platelets has been noted by other groups (3, 4, 21, 55). For example, a study examining the presence of histone proteins in platelets during sepsis showed that 5 to 35% of platelets were positive for the DNA-specific marker, DRAQ5 (55), and the presence of megakaryocyte-derived DNA in platelets was recently corroborated by an independent group (21). In this work, we have performed a detailed interrogation of pDNA content, demonstrating a biological role for platelets that was previously unappreciated: as scavengers of endogenous extracellular cfDNA, including tumor-derived and free fetal DNA.

We show that platelet uptake of DNA can occur by multiple mechanisms: endocytosis of DNA-loaded EVs from apoptotic cells and uptake of membrane-free DNA fragments. Previous reports indicated that DNA can bind to the platelet surface (56, 57). We found that DNA was also internalized by platelets and held in membrane-bound vesicles, where it was protected from exogenous DNase degradation and subsequently released upon platelet activation. DNA release could be prevented by using pharmacological inhibitors, thereby preserving the DNA within platelets where it is protected from degradation, increasing the yield of DNA obtained from platelet pellets.

Several physiological mechanisms exist to limit the abundance of cfDNA, as excess extrachromosomal DNA in the cytosol and plasma is proinflammatory (58, 59). We demonstrated higher and more variable levels of cfDNA in plasma in the setting of severe thrombocytopenia in both a mouse model and in patients. This supports a role for platelets in regulating the cfDNA content of plasma, although we cannot exclude that the higher level of cfDNA observed was due to other consequences of antibody-mediated platelet clearance.

Further work is required to ascertain whether platelets deploy their DNA content at sites of activation. An attractive hypothesis is that platelet clearance may control the nonspecific immunostimulatory activity of cfDNA in plasma, and that platelets may release DNA cargo to evoke an immune response in a targeted fashion at sites of platelet activation. Platelets have also been shown to mediate horizontal transfer of RNAs and transcription factors (60), either through uptake of



**Fig. 5. Detection of driver mutations in pDNA and cfDNA from mice with localized and metastatic colorectal adenocarcinoma and patients with premalignant colonic lesions.** (A) Schematic showing isolation of pDNA and cfDNA from C57BL/6 mice expressing  $KRAS^{G12D}$  and  $TP53$  (KP) mutations through the villin promoter resulting in locally invasive colorectal adenocarcinoma as well as mice with  $KRAS^{G12D}$ ,  $TP53$ , and  $NOTCH$  (KPN) mutations with aggressive, metastatic disease. (B) Waterfall plot showing fold difference in copies of  $KRAS^{G12D}$  detected using ddPCR per microliter of DNA extracted from the plasma (cfDNA) and platelets (pDNA) of KP and KPN mice, two weeks after tumor cell implantation.  $n = 5$  KP and 15 KPN mice. (C)  $KRAS^{G12D}$  copies detected by ddPCR, shown as copies per microliter of DNA extracted from plasma (cfDNA) and platelets (pDNA) of KP, KPN (circle), and KPC (square) mice compared with the total number of copies from plasma and platelets.  $n = 5$  KP, 15 KPN, and 6 KPC mice; datapoints

correspond to individual mice; mean  $\pm$  SEM is shown.  $**P < 0.01$ , as determined by a paired *t* test. (D and E) Representative ddPCR plots showing *KRAS*<sup>G12D</sup> (blue) in higher abundance in pDNA (top) than in cfDNA (bottom) in two representative mice, (D) a KPN mouse and (E) a KP mouse. (F) Pie charts showing *BRAF*<sup>V600E</sup> detection in high-risk colonic lesion patients (SSL, *n* = 31) and colonoscopy-screened controls (*n* = 14). Mutant *BRAF* (more than three positive events) was found in 16.1% (5/31) of SSL patients and 0% (0/14) of controls. (G) Relative *BRAF*<sup>V600E</sup> copy number in pDNA versus cfDNA for five SSL patients with detectable mutant *BRAF*. Data are log<sub>2</sub>-transformed with a pseudo count of 1. (H) ddPCR plot showing *BRAF*<sup>V600E</sup> (blue) in pDNA (left) and cfDNA (right) for two representative SSL patients. Data are combined.

whole platelets by phagocytes, endothelial cells, and smooth muscle cells or by the release of platelet-derived microvesicles (59). Whether platelets can similarly act as a vector for the horizontal transfer of DNA fragments, including potentially extrachromosomal DNA (59), and whether platelets modify sequestered DNA prior to release remain unknown.

Detection and analysis of cfDNA is rapidly being implemented in several clinical settings, including cancer screening, molecular profiling, and detection of residual disease as well as in prenatal diagnosis (61–63). Currently, a major limitation is the poor sensitivity for low-tumor burden disease. Preanalytical approaches to increase capture of ctDNA would be of interest. Anucleate RBCs have also been shown to contain DNA fragments, including cancer cell-derived DNA (64). In our studies, we did not detect *SRY* in red cell pellets of pregnant women, and a substantially lower ratio of mutant-to-wild-type alleles was observed in red cells than platelets in our coculture experiments, suggesting a superiority of platelets over red cells for liquid biopsy-based screening approaches. Our data suggest that a substantial proportion of ctDNA is contained within platelets and that this information is currently missed from standard sampling protocols that extract cfDNA from platelet-depleted plasma. This study paves the way for validation in larger clinical cohorts to test whether combining cfDNA with pDNA adds value as well as for future research to further establish a role for platelets in cfDNA homeostasis.

## Materials and methods

### Ethical approval, banking, and processing of samples

All procedures were performed in accordance with the ethical standards of the Declaration of Helsinki. Patient samples were collected by the Oxford Translational Gastroenterology Unit (TGU) and the Oxford Radcliffe Biobank (ORB). All patients provided informed consent, and procedures were approved by Yorkshire & The Humber - Sheffield Research Ethics Committee (REC reference number: 16/YH/0247), ORB research tissue bank ethics (REC reference 19/SC/0173) and Oxford GI Cohort ethics (REC reference 21/YH/0206).

Ten to 20 ml of peripheral blood was drawn into EDTA-coated (Fisher Scientific, cat # 367839) or cfDNA (Streck, cat # 218997) blood collection tubes from patients presenting with colorectal, esophageal, or pancreatic cancer. Ten ml of peripheral blood was collected into Streck tubes, from patients with sessile serrated lesions and from those on the bowel cancer screening pathway or those with inflammatory bowel disease undergoing colonoscopy.

For healthy controls, fully anonymised blood samples were collected from the Oxford Biomedical Research Laboratory under The INForMeD Study (REC reference 16/LO/1376). All donors provided informed consent, and blood was drawn into EDTA-coated tubes. Healthy donor platelet apheresis samples were provided by Oxford NHS Blood and Transplant (NHSBT) through the Non-Clinical Issue (NCI) service.

The usage and collection of healthy human donor blood samples in Edinburgh was authorised under the project “21-EMREC-041 – The Role of Inflammation in Human Immunity”. Freshly drawn blood from healthy human donors was collected into 50 ml BD Falcon tubes containing 4 ml of CPD buffer (16 mM citric acid, 90 mM sodium citrate, 16 mM NaH<sub>2</sub>PO<sub>4</sub>, 142 mM dextrose, pH 7.4).

For obstetric samples, 10 ml of blood was collected from mothers at the time of delivery, and postnatally at 24 and 48 hours after delivery into Streck blood collection tubes. Samples were taken at

the Department of Women's and Reproductive Health at the John Radcliffe Hospital Oxford and approved by the Oxfordshire Research Ethics Committee (07/H0607/74). Sex of the neonate was confirmed after delivery and all mothers provided written informed consent.

Patients with immune thrombocytopenic purpura (ITP) were consented under the INForMeD Study (REC reference 16/LO/1376) and 10 ml of peripheral blood was collected into Streck tubes.

### Animal models

All mice were bred and maintained in accordance with UK Home Office regulations. All experiments were performed under Project Licenses P2FF90EE8, POB63BC4D and PP6460882 and approved by the University of Oxford Animal Welfare and Ethical Review Body. All mice were housed in individually ventilated cages at the animal unit either at Functional Genetics Facility (Wellcome Centre for Human Genetics, University of Oxford) or the Biomedical Services Facility at the John Radcliffe Hospital (The MRC Weatherall Institute of Molecular Medicine, University of Oxford), in a specific-pathogen-free (SPF) facility, with unrestricted access to food and water, and were not involved in any previous procedures.

For the immune thrombocytopenia model, CD45.2 female, wild-type mice were injected intra-venously with either an anti-platelet antibody to reduce platelet count (Emfret, cat # R300) or an IgG control that had no cytotoxic effects on platelets in mice (Emfret, cat # C301). Mice were humanely culled and blood collected via cardiac puncture into EDTA coated microvettes (Sarstedt, cat # 20.1288) at 24, 72 and 120 hours post treatment. Platelet counts were determined using an automated blood cell counter. cfDNA was extracted from plasma and quantified.

For the CRC model, murine blood was drawn from KPN (*villinCre*<sup>ER</sup> *Kras*<sup>G12D/+</sup> *Trp53*<sup>fl/fl</sup> *R26*<sup>Nluc/+</sup>) and KP (*villinCre*<sup>ER</sup> *Kras*<sup>G12D/+</sup> *Trp53*<sup>fl/fl</sup>) mice via cardiac puncture into EDTA coated microvettes (Sarstedt, cat # 20.1288). All strains used in this model were maintained on C57BL/6J background. Both male and female KP and KPN mice were used.

For the orthotopic pancreatic cancer model, a total of 500 KPC-F cells (passages 9–11) were injected in a volume of 10  $\mu$ l in a mixture of 50% Matrigel and 50% DMEM (Gibco) using a 0.5 ml Micro-fine insulin syringe (BD). Following anesthesia of the mice using 4% isoflurane/oxygen and clipping of hair, an incision of 1 cm was made in the left abdominal site. The spleen and pancreas were exteriorized using forceps and KPC-F cells were injected in the tail of the pancreas. Successful injection was verified by the appearance of a wheal at the injection site with no leakage through the pancreatic capsule. Spleen and pancreas were gently moved back into the peritoneal cavity; the abdominal wall was sutured with absorbable 4.0 Vicryl suture (Ethicon) and the skin was closed using a 7 mm wound clip. Two weeks post-surgery, after the mice had recovered, blood was drawn via cardiac puncture into EDTA-coated microtainer tubes (BD).

For the cohort of KPC mice treated with aspirin, aspirin (NSC 27223; Selleck Chemicals, cat #. S3017) was administered via intraperitoneal injection on alternate days at a dose of 10 mg/kg. Treatment began on day 4 after tumor cell implantation and continued every other day until day 14. Control animals received intraperitoneal injections of vehicle (2% DMSO in PBS) on the same schedule.

### Blood cell isolation and nucleic acid extraction

Blood tubes were kept at room temperature and handled gently to avoid platelet activation. For human samples, platelets were isolated within 4 or 24 hours of blood sampling (depending on blood tube) using a modified previously published protocol (3). Briefly, whole

blood was supplemented with citrate-dextrose solution (ACD) (Sigma Aldrich, cat # C3821-50ML) and centrifuged at  $180 \times g$  for 20 min at room temperature. Eighty % of the supernatant was removed using a Pasteur pipette, diluted with platelet wash buffer and centrifuged at  $600 \times g$  for 20 min at room temperature. The resulting platelet pellet was resuspended in pre-warmed HEPES-modified Tyrode's buffer. The remaining platelet poor plasma was centrifuged at  $16'000 \times g$  for 10 min at  $4^{\circ}\text{C}$  to ensure the pelleting of all cellular components and the isolation of pure plasma cfDNA. For the collection of platelets from healthy donor platelet apheresis bags, samples were pelleted at  $600 \times g$  for 20 min at room temperature. Platelet numbers were determined using an automated blood cell counter (Horiba, Pentra ES 60 Cell Counter).

Human MNCs were isolated from freshly drawn venous blood into EDTA coated tubes. Whole blood was supplemented with ACD and centrifuged at  $180 \times g$  for 20 min at room temperature. The supernatant was removed and the remaining layer diluted with wash buffer (PBS pH 7.4 0.5% FBS, 2mM EDTA). Diluted blood was layered onto density gradient media (GE Healthcare, cat # 5442-03) and centrifuged for 20 min at  $600 \times g$ , low brake. MNCs were collected from the white inter-phase 'buffy coat' using a Pasteur pipette and transferred to a Falcon tube. Human RBCs were sampled from the upper layer of the RBC/granulocyte pellet and diluted with wash buffer. The remaining RBC/granulocyte pellet was resuspended with chilled RBC Lysis Buffer (eBioscience, cat # 00-4300-54) and incubated at  $4^{\circ}\text{C}$  for 5 min. The lysed sample was centrifuged at  $1000 \times g$  for 5 min, the supernatant discarded and the granulocyte pellet diluted with wash buffer. The remaining white granulocyte pellet was washed, centrifuged and stored at  $-80^{\circ}\text{C}$  until required. Purity studies using an automated cell counter provided no evidence of MNC or RBC contamination in the granulocyte pellet.

DNA was extracted from equal volumes of fresh platelets (resuspended in modified Tyrode's buffer) and 'platelet poor plasma' using the QIAamp cfDNA kit (Qiagen, cat # 55114), as per the manufacturer's instructions. Genomic DNA was extracted from nucleated cells using the DNeasy Blood & Tissue Kit (Qiagen, cat # 51104). Extracted DNA was quantified using a Qubit fluorometer (ThermoFisher Scientific) and stored at  $-80^{\circ}\text{C}$  until required.

Mouse platelets were isolated within four hours of blood collection via cardiac puncture. Whole blood was supplemented with ACD and centrifuged at  $100 \times g$  for 10 min at room temperature. The plasma was removed and collected into a 1.5 ml Eppendorf tube. The plasma was then diluted with platelet wash buffer and centrifuged at  $3500 \times g$  for 11 min to pellet the platelets. The platelet pellet was resuspended in pre-warmed HEPES-modified Tyrode's buffer. The platelet poor plasma was removed and centrifuged at  $1300$  to  $16'000 \times g$  for 10 min at  $4^{\circ}\text{C}$  to remove all cells and debris, and the final supernatant collected for cfDNA isolation.

### Cell lines

The following cancer cell lines were used as positive and negative controls for common cancer-associated mutations; HCT116 (CVCL S744) *PIK3CA*<sup>H1047R</sup>, LS180 (CVCL 0397) *KRAS*<sup>G12D</sup>, COLO205 (CVCL 0218) *BRAF*<sup>V600E</sup>, HEL (CVCL\_0001) *JAK2*<sup>V617F</sup>. All cell lines were purchased from American Type Culture Collection (cat # CCL-247, CL-187, CCL-222 and TIB-180 for HCT116, LS180, COLO205 and HEL cells respectively). All cells were cultured according to American Type Culture Collection recommendations and tested monthly for mycoplasma contamination.

NKPCB-NBC12.1F (KPC-F) cells, derived from a KPC mouse (*LSL-Kras*<sup>G12D</sup>; *LSL-Trp53*<sup>R172H</sup>; *Pdx1Cre*) were cultured in DMEM (Gibco) supplemented with 10% FBS, 2 mM glutamine, 100 U/ml penicillin, 100 µg/ml streptomycin and maintained at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$ .

The human Burkitt's lymphoma cell line, BL2 [CVCL 1966, cat # CRL-2959] was used to generate tumor cell-derived EVs. Both standard BL2 cells and BL2 cells stably transfected with the apoptosis-suppressing

gene *BCL2* were used, to obtain apo EVs from BL2 cells and nonapo EVs from apoptosis-resistant BL2-Bcl-2 cells. Protein expression was regularly tested by flow cytometry, confirming that 98% of the BL2-Bcl-2 cells expressed BCL2. Both cell lines were maintained in 50% X-VIVO medium (50% Gibco RPMI-1640, 50% X-VIVO-20 medium; Lonza, Basel, Switzerland), supplemented with 50 U/ml penicillin and 50 µg/ml streptomycin at  $37^{\circ}\text{C}$  with 5%  $\text{CO}_2$ .

### Generation of EVs from BL2 and BL2-Bcl-2 cells

BL2 and BL2-Bcl-2 cells were cultured at  $20 \times 10^6/\text{ml}$  in  $0.1 \mu\text{m}$  filtered 50% X-vivo 20 and irradiated with ultraviolet light to induce apoptosis with 6 doses at  $50\text{mJ}/\text{cm}^2$ , for a total of  $300\text{mJ}/\text{cm}^2$ . Apoptosis was monitored hourly by Annexin V and Sytox Blue staining. To isolate EVs, the culture supernatant was centrifuged at  $25 \times g$  for 1 hour followed by sequential filtering through a  $5 \mu\text{m}$  mesh filter and a  $1.2 \mu\text{m}$  syringe filter.

### Purification and labeling of apo and nonapo EVs

The EVs were purified from soluble factors and proteins from the cell culture by size exclusion chromatography using in-house prepared sepharose columns (ThermoFisher, cat # 45-000-067 and Sigma Aldrich, cat # CL6B200-100ML). After purification, EV concentration and size profile was measured by Nanoparticle Tracking Analysis using a NanoSight LM10 and Malvern NanoSight software (v3.3). The software analyzed each sample five times to give the average concentration, mean and modal size of each sample. EV size ranged between 150-200 nm and the concentration was between  $100 \times 10^8$  to  $200 \times 10^8/\text{ml}$ . The number of EVs added to platelets was standardized across experiments, using a ratio of  $50 \times 10^8$  EVs with  $1 \times 10^7$  healthy donor platelets. For imaging experiments, prior to incubation with platelets, the purified EVs were resuspended in  $0.1 \mu\text{m}$  filtered HBSS and were stained with Biotium CF568 NHS ester with a covalent attachment of the dye to EV proteins, according to the manufacturer's protocol (Biotium, cat # NC1542764). To remove unbound dye, the EVs were purified again by size exclusion chromatography through sepharose columns.

### Flow cytometry and fluorescence-activated cell sorting (FACS)

All experiments were carried out using an LSRFortessa X-20 flow cytometer (Becton, Dickinson and Company). Platelets and paired plasma were prepared as previously described, pelleted and resuspended in HEPES-modified Tyrode's buffer. The samples were stained with a cocktail of antibodies for 20 min at room temperature in the dark, prior to being washed and resuspended in fresh buffer for flow cytometric analysis. The antibodies used are as follows, CD42 FITC, CD41 PE-Cy7, CD3 PE, CD11b and CD33 APC, CD19 APC-Cy7 and CD235a BV650. To assess the nucleic acid content of platelets, healthy donor blood was drawn into EDTA-coated tubes and platelets were isolated as previously described. The platelets were stained with the antibody CD42 PE-Cy7 and nucleic acid stains DRAQ5 (DNA) and Syto-13 (RNA) for 20 min at room temperature in the dark. After incubation, the platelets were washed and resuspended in fresh HEPES-modified Tyrode's buffer for analysis. Compensation beads (were stained and analyzed in parallel, as an additional negative control to fluorescence minus one (FMO) controls to confirm no non-specific binding of the nucleic acid stains (BD Invitrogen, cat # 01-1111-42). To assess platelet activation, platelets were isolated from healthy donor blood drawn into citrate tubes. Following isolation, platelets were maintained at a physiological concentration of  $\sim 300 \times 10^6/\text{ml}$ .

To evaluate the activation status and DNA content of healthy donor platelets, after isolation, platelets were stained with CD42 FITC, and antibodies for activation; CD63 PB and CD62P BV857. The DNA content was simultaneously assessed by the inclusion of the DNA marker, DRAQ5. After a 20-min incubation at room temperature, the platelets were washed and resuspended in HEPES-modified Tyrode's buffer.

To determine changes in pDNA content upon activation,  $10 \mu\text{l}$  of platelets were added to Eppendorf tubes containing the antibodies

CD42 FITC, CD63 PB and CD62P BV857 and the DNA stain DRAQ5. The platelets were then treated with 3  $\mu$ l of either a 'resting' or an 'activation' buffer, both comprising HEPES-modified Tyrode's buffer supplemented with 20 mM calcium (Sigma-Aldrich, cat. # 21115-100ML). For the activation buffer, the agonist TRAP-6 was added at concentrations of 5, 10, 50, or 100  $\mu$ M (10x the final concentration, Bachem AG, cat. # 4031274.0025). After a 15-min incubation at room temperature, the platelets were washed twice prior to resuspension in HEPES-modified Tyrode's buffer and analyzed.

The BD FACSymphony S6 was used for cell sorting. 30  $\mu$ l of healthy donor blood was diluted with an antibody cocktail to a final volume of 100  $\mu$ l. The platelets were incubated for 20 min at room temperature in the dark, washed and resuspended in fresh buffer. The staining solution included the following antibodies, CD42 FITC, CD3 BV421, CD11b and CD33 APC, CD19 APC-Cy7 and CD235a FITC. Platelets (CD42<sup>+</sup>, CD3<sup>+</sup>, CD11b<sup>+</sup>, CD33<sup>+</sup>, CD19<sup>+</sup> and CD235a<sup>+</sup>) were sorted into 15 ml falcon tubes containing HEPES-modified Tyrode's buffer. The platelets were centrifuged at 600  $\times$  g for 20 min, the supernatant removed, and pellet frozen at -20°C.

Single color-stained controls and FMO controls were used for all experiments. FACS data were collected using BD FACSDiva software (v9.0) and gates were set using FMO controls and negative populations. All antibodies and nucleic acid stains are listed in table S7.

### Droplet digital PCR

Droplet digital PCR (ddPCR) assays were ordered from Bio-Rad. All ddPCR assays were conducted on a QX200 Droplet Digital PCR System using the manufacturer's recommended protocol and reagents (Bio-Rad Laboratories). Positive and negative controls were included in every assay. Analysis was performed using the QuantaSoft software (Bio-Rad Laboratories, Watford UK).

To detect mouse *KRAS*<sup>G12D</sup>, a ddPCR assay was designed for the detection of the *KRAS* p.G12D c.35G>A point mutation. Primers for the detection of *KRAS* were as follows: forward 5'-GCCTGCTGAAATGACTGAG-3' and reverse 5'-CGTAGGGTCATACTCATCCAC-3'. Two dual labeled probes were also used to target the wild-type and mutant sequences.

Wild-type 5'-HEX-ACGCC[+A][+C]CAG[+C]TCCAA-BHQ1-3' and mutant 5'-6FAM-AC[+G]CC[+A][+T][+C]AG[+C]TCCAA-BHQ1-3'. Square brackets indicate LNA bases (Merck Life Sciences).

### Synthetic DNA oligo generation

Single stranded DNA oligos were designed to include the mouse *KRAS*<sup>G12D</sup> genetic marker and synthesized by Integrated DNA Technologies (IDT, Iowa USA). Synthesized DNA was reconstituted according to the manufacturers protocol. PCR assays for the detection of *KRAS* were performed using the KAPA2G Fast HotStart PCR Kit (Roche, cat # KK5512) and primers were as follows: forward 5'-GCCTGCTGAAATGACTGAG-3' and reverse 5'-CGTAGGGTCATACTCATCCAC-3'. Fragmentation profiles of DNA oligos were analyzed using an Agilent Tapestation 2200 system (Agilent Technologies). Electrophoresis data was analyzed using the Tapestation Analysis Software (Agilent).

### In vitro coculture of platelets, tumor cells, EVs, and synthetic DNA oligos

To explore uptake of DNA shed from tumor cells, platelets (3  $\times$  10<sup>5</sup>/μl), MNCs (0.075  $\times$  10<sup>5</sup>/μl) and RBCs (3  $\times$  10<sup>5</sup>/μl) from healthy donors were added to individual wells within a 24-well plate and a 1.0  $\mu$ m TC Insert (Sartstedt, cat # 83.3932.101) placed into the well. Cancer cells (0.03  $\times$  10<sup>5</sup>/μl) were carefully placed in the inserts and the plate incubated for approximately eight hours at 37°C. Control samples were incubated simultaneously but without the addition of tumor cells. After incubation, the inserts were removed and the blood cells transferred to Eppendorfs. To ensure complete removal of any contaminating cfDNA in the media, all cells/platelets were washed and centrifuged at 600  $\times$  g for 10 min. The supernatant was removed and the pelleted blood cells re-suspended in fresh buffer. The cells were washed for a total of three times and

finally resuspended in 1 ml of fresh pre-warmed HEPES-modified Tyrode's buffer prior to DNA extraction using the QIAamp cfDNA kit. Following DNA extraction, the samples were quantified and ddPCR was used to identify the presence of mutant alleles.

To investigate platelet uptake of DNA-containing EVs, 50  $\times$  10<sup>8</sup> labeled, purified apo and nonapo EVs resuspended in Tyrode's buffer were incubated with 0.1  $\times$  10<sup>8</sup> platelets for three hours at 37°C with gentle inversion every 30 min. Following incubation, the samples were centrifuged for 20 min at 800  $\times$  g. The supernatant was discarded, and the platelets resuspended in 300  $\mu$ l of PBS. EV and platelet pellets were treated with DNase as per manufacturer's protocol (Sigma Aldrich, cat # AMPD1) and then DNA extracted using the QIAamp cfDNA kit. Following DNA extraction, the samples were quantified and assayed for the male *SRY* gene using ddPCR.

To understand whether platelets also take up "free" DNA fragments, platelets were incubated with synthetic DNA fragments ranging from 128 to 657 bp. Healthy donor human platelets were aliquoted into Eppendorf tubes and spiked with reconstituted synthetic DNA oligos for a final concentration of 10 ng/ml. The platelets were mixed gently by pipetting and incubated at 37°C overnight. Following incubation, the platelets were centrifuged for 10 min at 600  $\times$  g. The supernatant (containing excess DNA oligo) was discarded, and the platelets resuspended in fresh buffer. The platelets were washed for a total of two times and finally resuspended in 1ml of fresh pre-warmed HEPES-modified Tyrode's buffer prior to DNA extraction using the QIAamp cfDNA kit. Eluted DNA was PCR amplified using the KAPA2G Fast HotStart PCR Kit (Roche, cat # KK5512) and fragment length was confirmed using the Agilent Tapestation.

### DNase treatment to investigate pDNA internalization

Platelets were incubated with HEL erythroleukemia cells as previously described. Following incubation, platelets were treated with DNase according to the manufacturers protocol (Ambion DNA-free Kit, cat # AM1906). To confirm catalytic activity of the enzyme, DNase was also added to conditioned media. Following DNase treatment, the platelet samples were washed with 500  $\mu$ l of 5mM EDTA and centrifuged at 600  $\times$  g for 10 min. The supernatant was removed and the wash repeated. After the second centrifugation, the supernatant was discarded and the platelet pellet resuspended in 1 ml of fresh pre-warmed HEPES-modified Tyrode's buffer. Following DNase inactivation of the conditioned media, 900  $\mu$ l of HEPES buffer was added to a final volume of 1 ml in preparation for DNA extraction. For control samples, no DNase was added and 10  $\mu$ l of 1M tris-HCL was added in replacement of 10X DNase I Buffer for the platelet samples.

In a separate experiment, healthy donor platelets or an equal volume of HEPES-modified Tyrode's buffer (to mimic plasma) was spiked with a 260 bp synthetic DNA oligo and incubated overnight. The samples were then treated with DNase according to the manufacturers protocol (Life technologies, RapidOut DNA Removal Kit, cat # K2981). Following DNase treatment, the platelet samples were washed and centrifuged at 600  $\times$  g for 10 min. The supernatant was removed and platelet pellet wash repeated. After the second centrifugation, the supernatant was discarded and the platelet pellet resuspended in 1 ml of fresh pre-warmed HEPES-modified Tyrode's buffer. For 'cfDNA' (buffer) samples, DNase was inactivated and 900  $\mu$ l of HEPES buffer was added to a final volume of 1 ml. DNA was extracted from all samples using the QIAamp cfDNA kit. Eluted DNA was PCR amplified and the fragment length confirmed as previously described. For control samples, no DNase was added and 10  $\mu$ l of 1M tris-HCL was added in replacement of 10X DNase I Buffer for the platelet samples.

### Inhibition of cargo trafficking by platelets

Healthy donor human platelets were treated with the following inhibitors or appropriate negative control molecules; Dynole 34-2 (Abcam, cat # ab120463, 30  $\mu$ M), Dynole 31-2, Negative control (Abcam, cat #

ab120464-5mg, 30  $\mu$ M), Pitstop (Abcam, cat # ab120687, 25  $\mu$ M), Pitstop 2 Negative control, (Abcam, cat # ab120688, 25  $\mu$ M), Cytochalasin D (Santa Cruz Biotechnology, cat # CAS 22144-77-0, 5  $\mu$ M), Nystatin (Merck Life Science UK, cat # N9150-20ML, 30 units/ml), Amiloride (Abcam cat # ab120281, 500  $\mu$ M), Imipramine (Abcam, cat # ab141046, 5  $\mu$ M). After treatment, platelets were spiked with a 260 bp synthetic DNA oligo for a final concentration of 10 ng/ml and incubated at 37°C overnight. Following incubation, the platelets were centrifuged for 10 min at 600  $\times$  g. The supernatant (containing excess inhibitor/negative control and DNA oligo) was discarded, and the platelets re-suspended in fresh buffer. The platelets were washed for a total of two times and finally resuspended in 1 ml of fresh pre-warmed HEPES-modified Tyrode's buffer prior to DNA extraction using the QIAamp cfDNA kit. Eluted DNA was PCR amplified and the fragment length confirmed as previously described.

To assess the impact of platelet activation inhibition on pDNA uptake, platelets were treated with 500, 1000 or 2000  $\mu$ M aspirin (Merck Life Sciences, cat # A2093-500G) or 5, 50, 100 or 200  $\mu$ M ibrutinib (PCI-32765) (Stratex Scientific Ltd, cat # S2680-SEL-5mg). Following treatment, a 260 or 367 bp synthetic DNA oligo was added to resting platelets for a final concentration of 10 ng/ml DNA and incubated at 37°C overnight. Following incubation, the platelets were centrifuged for 10 min at 600  $\times$  g. The supernatant (containing excess inhibitor/negative control and DNA oligo) was discarded, and the platelets re-suspended in fresh buffer. The platelets were washed for a total of two times and finally resuspended in 1 ml of fresh pre-warmed HEPES-modified Tyrode's buffer prior to DNA extraction. Eluted DNA was amplified by PCR and the Agilent TapeStation was used to confirm fragment length.

To assess the impact of dynamin inhibition on platelet secretion, platelets were treated with Dynole or paired negative control, exposed to the 260 bp synthetic DNA oligo and incubated overnight. Following incubation, the platelets were pelleted, resuspended in fresh buffer, re-treated with the inhibitor or negative control and incubated for 3 hours at 37°C. After the second incubation, the platelets were pelleted and discarded, and the supernatant (platelet 'releasate') was collected. The supernatant was centrifuged at 16'000  $\times$  g for 5 min to ensure complete removal of any contaminating platelets and subject to DNA extraction, PCR amplification and fragment analysis.

To understand the impact of dynamin inhibition on platelet EV internalization,  $0.1 \times 10^8$  healthy donor platelets were incubated with either the dynamin inhibitor, paired negative control or DMSO for 30 min at 37°C. After incubation,  $50 \times 10^8$  labeled, purified apo EVs resuspended in Tyrode's buffer were added to the treated platelets and incubated as previously described prior to DNase treatment, DNA extraction and SRY ddPCR.

### H3K27ac ChIPmentation of platelet chromatin

Platelets were isolated from plateletpheresis samples from 3 independent healthy donors. 13 ml of each sample was centrifuged at 600  $\times$  g for 30 min at room temperature, the supernatant was removed and platelet pellets were immediately used for H3K27ac ChIPmentation using a protocol tailored for low-DNA input samples from a published protocol (65), using the H3K27ac antibody (Diagenode, cat # C15410196). In brief, bead-bound chromatin underwent a one-step adaptor insertion via Tn5 tagmentation and only one post-PCR DNA purification was performed to reduce sample loss.

For each reaction, 5  $\mu$ l A-Dynabeads (Invitrogen cat # 10001D) were coated with the H3K27ac antibody (Diagenode, cat # C15410196) (150  $\mu$ l PBS, 0.5% BSA, 1x Proteinase inhibitor). The antibody-bound beads were then washed in PBS and 0.5% FBS, before being added to samples for overnight incubation.

Platelet pellets were incubated in 600  $\mu$ l lysis buffer (50 mM Tris pH 8, 10 mM EDTA, 0.5% SDS, 1x Protease Inhibitor) for 20 min on ice. Then, 1% Triton-X100 was added, and the samples centrifuged to

pellet insoluble material. The supernatant was split into three 200  $\mu$ l sub-samples, and the samples were incubated overnight with the H3K27ac antibody coated beads. Before incubation, 5% of the sample was kept aside for input control tagmentation.

After overnight incubation, the beads were washed three times with RIPA wash buffer (50 mM HEPES pH 7.6, 500 mM LiCl, 1 mM EDTA, 1% NP-40 / IGEAL CA-630, 0.7% Na-Deoxycholate), followed by one wash with TE and then 10 mM Tris pH8. Beads and input control samples were then tagmented in tagmentation buffer (50 mM Tris pH 8, 25 mM MgCl<sub>2</sub>, 50% Dimethylformamide) with Tn5 transposase (Illumina, cat # 20034197) for 10 min at 37°C and immediately cooled down. Beads were immediately washed. Library preparation was then conducted with Nextera-based sequencing primers (66). qPCR and fragmentation analysis were performed.

### Immunofluorescence microscopy

**Live:** To visualize pDNA uptake in vitro, washed human platelets were stained with anti-CD42 AF488 (BioLegend, cat # 303914) for 30 min at room temperature. Separately, COLO205 cells or H2B-mCherry HEL cells were labeled using either the highly specific live cell DNA stain, SiR-DNA (Spirochrome, cat # CHF260.00), NUCLEAR-ID Red DNA stain or 4',6-diamidino-2-phenylindole (DAPI) (HEL only) as per the manufacturers' protocols (Enzo Life Sciences, cat # ENZ-52406). Stained COLO205 or H2B-mCherry HEL cells were then washed and added to collagen type-1 (Corning, cat # 354236)-coated microscopy slides (Greiner Bio-One, cat # 543079). CD42 stained platelets were added to the wells containing tumor cells immediately before imaging. To analyze endogenous wild-type DNA within healthy donor platelets, platelets were co-stained with anti-CD42 AF488 and NUCLEAR-ID Red DNA and imaged alone.

Regarding platelet uptake of tumor derived DNA from conditioned media, COLO205 cells were labeled with NUCLEAR-ID Red. Following staining, the cells were washed with MEM Alpha 5% FCS, pelleted and the supernatant removed. The cells were washed a further two times to ensure complete removal of the DNA stain and resuspended in 200  $\mu$ l of fresh media. The cells were incubated at 37°C and vortexed periodically to encourage cell death and DNA release. After two hours, the COLO205 cells were centrifuged at 16'000  $\times$  g for 10 min at 4°C. Supernatant was removed from the pelleted cells, transferred to a fresh Eppendorf and the cell pellet discarded. Healthy donor platelets were stained with anti-CD42 AF488 and added to wells of Poly-L-lysine coated slides (Ibidi, cat # 81201). DNA labeled COLO205 cell conditioned media was added to the platelets immediately before imaging. All live imaging was performed using the Zeiss Spinning Disk Confocal with a 63X oil immersion objective.

**Fixed:** BL2 cell-derived apo EVs were isolated, labeled with an amine-reactive fluorescent dye (Biotium, cat # 92131) and frozen as previously described. Prior to imaging, the EVs were thawed and co-stained with DAPI for 30 min at room temperature. The EVs were washed twice, pelleted at 20'000  $\times$  g for 30 min at 4°C and resuspended in HBSS buffer. Labeled EVs were then added to Poly-L-lysine coated chamber slides and incubated overnight at 4°C. Following incubation, the supernatant was carefully removed and the EVs fixed with 4% formaldehyde at room temperature for 30 min. The fixative was then removed and the EVs gently washed three times with PBS. To confirm imaging of EVs and not auto-fluorescent debris, PBS was stained with DAPI, incubated overnight, and fixed onto slides alongside the EV preparations. The slides were mounted and the EVs and PBS controls resolved by fluorescence microscopy using a Zeiss LSM900 with a 63X oil immersion objective.

To visualize platelet sequestration of EVs,  $5 \times 10^9$  apo EVs labeled with an amine-reactive fluorescent dye were thawed and added to 1 ml of  $10 \times 10^6$  washed healthy donor platelets and incubated for three hours at 37°C. To ensure adequate mixing, the cells were inverted

gently every 30 min. Following incubation, the cells were centrifuged at  $800 \times g$  for 20 min, low brake. The supernatant (containing the EVs) was removed, and the platelets resuspended in PBS. Platelets were centrifuged onto a Poly-L-lysine coated chamber slide at  $600 \times g$  for 20 min and fixed in 2% formaldehyde/PBS for 10 min at room temperature. The fixative was removed and the platelets washed twice with PBS. The platelets were then labeled with anti-CD42 AF488 for 20 min at room temperature. After labeling, the slides were washed, mounted with ProLong Gold Antifade Mountant (ThermoFisher, cat # P36930) and imaged using the Zeiss LSM900.

### Histology, immunohistochemistry, and imaging of peripheral blood films

Cell contamination was assessed after cytopspin of the resuspended platelet pellet onto a glass slide (5 min at 500 r.p.m.) and May–Grünwald Giemsa staining, according to standard protocols. Images were obtained using an Olympus BX60 Microscope.

KPC mice were sacrificed, and pancreatic tissue was taken for p53 immunohistochemical staining. Briefly, a total of 24 hours post fixation in 10% neutral buffered formalin at 4°C, tissues were transferred to 70% ethanol and stored overnight at 4°C. Fixed tissues were processed overnight using the STP120 Spin Tissue Processor (Thermo Fisher Scientific) and embedded the next day in paraffin wax. Tissues were cut into 4  $\mu$ m sections using a Leica RM215 microtome and adhered onto SuperFrost Ultra Plus Adhesion slides (Thermo Fisher Scientific) before drying overnight in a 37°C incubator. After deparaffinization, rehydration in an ethanol gradient (100%, 70%, 50%), and antigen retrieval with Citrate Buffer Antigen Retriever (Sigma-Aldrich), slides were incubated with Dual Endogenous Enzyme-Blocking Reagent (Dako) for 1 hour, washed in PBS, incubated with 5% goat serum for 30 min, washed in PBS again, and incubated with the primary antibody, p53 (Cell Signaling Technologies, cat # 2524) diluted 1:2000 for 2 hours at room temperature. Slides were then washed in PBS with 0.05% Polysorbate 20 and incubated with the secondary antibody, horse anti-mouse IgG (Cell Signaling Technologies, cat # 7076), diluted 1:100 for 45 min at room temperature, washed in PBS with 0.05% Polysorbate 20 again, and incubated with 3,3'-Diaminobenzidine (DAB) chromogen for 3 min at room temperature. After staining, slides were dehydrated in an ethanol gradient (50% 70%, 100%), followed by tissue clearing in xylene (ThermoFisher). Slides were mounted with dibutylphthalate polystyrene xylene (DPX; Sigma-Aldrich), dried for 24 hours and scanned using an Aperio ScanScope CS digital slide scanner (Leica Biosystems) at 20X magnification.

Healthy donor blood was used to prepare a peripheral blood film. The blood smear was fixed with 100% methanol for 30 s and stained with CD42 AF488 (BioLegend, cat # 303914) to identify platelets and the DNA stain Hoechst (Invitrogen, cat # H3570), to identify nucleated cells. The slide was mounted with a coverslip and imaged using the Leica SPE.

### FISH

Platelets in suspension were centrifuged onto Poly-L-lysine coated chamber slides and fixed in 2% formaldehyde/PBS for 10 min at room temperature. The platelets were then permeabilized in 0.5% Triton X-100/PBS for 10 min at room temperature and washed twice in 0.05% Triton-X 100/PBS. The platelets were washed in 0.02% Tween20/PBS (PBST) and incubated with the primary anti- $\beta$ -tubulin antibody (Sigma Aldrich, cat # T5201) diluted in blocking buffer for 30 min at 37°C in a humid box. Following primary labeling, the platelets were washed with PBST and incubated with the secondary donkey anti-mouse IgG H&L AF405 antibody (Abcam, cat # ab175658) diluted in blocking buffer for 30 min at 37°C. The labeled platelets were then washed with PBST, fixed and washed with PBS for 10 min at room temperature. FISH staining was carried out according to the manufacturer's protocol. Briefly, slides were incubated in 0.1N HCL, washed with 2x SSC and dehydrated in 70% ethanol for 3 min. Equal volumes of X- and

Y-chromosome probes (Metasystems, cat # D-0323-050-FI and D-0324-100-OR) were added to the slide and covered with a  $22 \times 22$  mm<sup>2</sup> coverslip. The slides were incubated at 85°C for 5 min, sealed with rubber cement and incubated in a humidified chamber at 37°C overnight. Following hybridization, the rubber seal was removed, and the slides washed in 0.1X SSC at 60°C. Slides were mounted and imaged using the Zeiss LSM900 with a 63X oil immersion objective. Control cells (HEL cells and MNCs) were fixed onto slides, permeabilized and labeled with the X- and Y- chromosome paints. Following overnight hybridization, the cells were washed and counterstained with DAPI.

### Airyscan imaging of platelet ultrastructure

Glass coverslips (VWR, cat # 631-0150) were coated with 5  $\mu$ g/ml fibrillar collagen-I (Horm), (Nycomed) and incubated at 4°C overnight in a 24 well plate. The coverslips were blocked with 5 mg/ml BSA for 1 hour at room temperature. After blocking, the BSA was removed and 300  $\mu$ l of washed healthy donor platelets diluted to  $1 \times 10^7$ /ml was added to the wells. The platelets were left to spread for 45 min in a humidified chamber at 37°C. Following incubation, the liquid was carefully removed and washed with PBS. The platelets were fixed with 5% formalin for 10 min at room temperature, washed with PBS and permeabilized with 0.1% Triton-X 100/PBS (Sigma, cat # SLCF5969) for 5 min. The slides were washed with PBS and blocked for 30 min with blocking buffer (1% BSA + 2% goat + 2% donkey serum). The primary antibodies: mouse anti- human CD63, mouse anti-human LAMP2, rabbit anti- human TOM20, rabbit anti- human CD81 and mouse anti-human CD62P were diluted in blocking buffer and incubated with the platelets at 4°C overnight. Following primary incubation, the coverslips were washed three times with PBS and incubated with the DNA stains (DRAQ5, NUCLEAR-ID Red, Hoechst) and the secondary antibodies; goat anti- mouse IgG H&L AF568 and goat anti- rabbit IgG H&L AF488 for 45 min at 37°C. Following incubation, the coverslips were washed and mounted onto imaging slides using Hydromount (Scientific Laboratory Supplies Ltd, cat # NAT1324). The slides were imaged using the Airyscan function of a Zeiss LSM 980 confocal microscope (63X objective lens, NA 1.4).

### Quantitative analysis of imaging data and image analysis software

Images were analyzed using ImageJ (v2.1.0) (National Institute of Health, US-MD) and 3D reconstructions were generated in Imaris Viewer v9.7.0 (Oxford Instruments, Abingdon). CellProfiler (v4.0.7) (Broad Institute, US-MA) was used to quantitatively analyze platelet-tumor cell interaction. In brief, cells were segmented using the 'Identify Primary/Secondary Object' module. Thresholding strategies were employed to accurately mask cells for segmentation. Platelets were tracked using the 'Track Objects' module. Finally, AF647 intensity was measured using 'Measure Object Intensity' and data 'Exported to Spreadsheet'. This image-analysis pipeline, as well as others can be recreated in Cell Profiler tool (freely available from the Broad Institute at [www.cellprofiler.org](http://www.cellprofiler.org)).

For colocalization analysis, we calculated the Pearson's correlation coefficient,  $r$  and Mander's overlap coefficient,  $R$  for each platelet by manually defining a cuboid region ( $90 \times 90$  pixels;  $3.8 \times 3.8$   $\mu$ m), enclosing fluorescence from all three channels. The equations used to calculate fluorescence correlation coefficients are described in the Supplementary Text, with calculations performed using MATLAB as described by Jung *et al.* (67).

To assess the location of the DNA fluorescence within the platelet we developed a simple algorithm in MATLAB to calculate the total intensity of the DNA marker in a three-dimensional (3D) platelet mask that was subsequently eroded away from the surface to the center. First, we generated a 3D platelet mask by applying a one level Otsu thresholding algorithm to the CD62P fluorescence signal. The DNA intensity within the platelet was obtained by integrating the DRAQ5 fluorescence signal from pixels within the 3D platelet mask. To determine

the position of the DNA fluorescence within the platelet, we used a series of one pixel erosion operations on the original 3D mask. After each erosion step, a convex hull was applied to ensure there were no 'holes' in the mask. The integrated DNA fluorescence intensity was then obtained within this new eroded mask. This procedure was repeated until a single pixel of the 3D platelet mask remained i.e., the center of the platelet had been reached.

### Whole-genome sequencing (WGS)

WGS was performed on paired pDNA and cfDNA isolated simultaneously from the same peripheral blood samples of 15 patients with a recent diagnosis of gastrointestinal adenocarcinoma. Libraries were prepared from samples that were confirmed (using the Agilent TapeStation) to have both short DNA fragments of approximately 160 bp in addition to longer peaks of ~12'000 bp. l-pDNA was fragmented prior to library preparation. Briefly, 15 µl of DNA diluted in TE buffer (1mM Tris-HCl, pH 8.0, 0.1 mM EDTA) was added to a Covaris microtube-15 (Covaris, cat # 520145) and briefly centrifuged at 3000 × g for 1 min. The microtube was loaded onto the Covaris ME220 Focused-ultrasonicator and the protocol run using the treatment conditions required for a 150 bp target peak. Fragment size was determined using the Agilent TapeStation 2200 system (Agilent Technologies) and the sonication was repeated if DNA fragments were > 150 bp.

DNA-Seq libraries were prepared using the NEBNext Ultra II DNA Sample Preparation Kit for Illumina (New England BioLabs, Ipswich, MA, USA) according to the manufacturer's protocol. pDNA was size selected into 2 groups: short fragments (s-pDNA; under 600 bp) and long fragments (l-pDNA; over 600 bp). The short fragments were further cleaned to remove <100 bp fragments and large platelet fragments were fragmented via sonication as previously described. Following end repair and adapter ligation, adapter-ligated DNA fragments were amplified in enrichment PCR to generate final libraries. Cleaned up libraries were then multiplexed, and 1.5 pM libraries analyzed on an Illumina NextSeq 500 (300 cycle PE) at low-pass (0.1X) for all samples, and 10X for the cfDNA and s-pDNA, using four lanes in each sample with 4% PhiX to monitor sequencing performance. This amounted to 240 fastq files.

### Data preprocessing

Quality control of all fastq files was conducted by running FastQC (v0.11.8) (68). Reads were adaptor- and quality-trimmed using Trim Galore! (69) (v0.6.5) and aligned to the GATK Genome Reference Consortium Human Build 38 (GRCh38) (70) using bwa-mem (v0.7.17) (71). Reads were aligned to alternate contigs to represent common complex variation using bwa\_postalt.js (<https://github.com/lh3/bwa/blob/master/bwakit/README.md>) (71). Resulting BAM files were merged, sorted and indexed with samtools (v1.13.0) (72).

GATK toolkit (v4.1.7.0) was used to mark optical and PCR duplicates, estimate library complexity, and calculate summary metrics on insert sizes (73). Mapping rate was evaluated with samtools flagstat and the number of reads mapped to each chromosome and the mitochondrial genome with idxstats. Repetitive elements in the ENCODE blacklist (74) were excluded using bedtools intersect (v2.30.0) (75).

### Insert size distribution

For all uniquely mapped, nonduplicated, and properly paired read pairs, the frequency of insert size values was counted across all autosomes and chromosome X using samtools (72) and R. The distributions were plotted using ggplot2 (76).

### Read depth around transcription start sites

Transcription start sites were defined by refTSS (v3.3) (77). We used the nearest refTSS entry to the 5' coding sequence of all protein coding genes. Coverage per base in a 2 kilobase pair (kbp) region around each TSS was extracted with Rsamtools, counting only uniquely mapped,

non-duplicate, correctly paired reads with insert sizes between 150 and 170 bp ("mono-nucleosome") or between 280 and 350 bp ("di-nucleosome"). Peripheral blood mononuclear cell (PBMC) gene expression values were taken from Blueprint Epigenome experiment: EGAX00001327129 (<http://dcc.blueprint-epigenome.eu/#/experiments/ERX1123729>).

Briefly, genes with posterior mean estimates of transcripts per million > 15 were defined as "highly expressed" in PBMCs.

### Dinucleosome peak difference quantification

A  $\chi^2$  test of independence was performed in R with a simulated p-value to examine the relation between DNA type (cfDNA and s-pDNA) and the proportion of di-nucleosome fragments. We separated the fragments in (i) fragments below 250 bp representing mono-nucleosome fragments and (ii) fragments above 250 bp representing di-nucleosome fragments. The proportion of di-nucleosome fragments was higher in s-pDNA than in cfDNA ( $P < 0.0005$  in all samples).

### Chromosome coverage

Coverage was calculated using samtools bedcov, using a bed file of 10 kbp non-overlapping windows for all main chromosomes, but excluding reads with mapQ < 1. Position with mappability (as defined by umap: <https://bismap.hoffmanlab.org/>) in the lowest 8 percentile were removed (78). The Coverage was then corrected for mappability and GC content bias using a linear regression model and visualized with karyoplotR (79).

### Gini index

After filtering for reads that were in pair, uniquely mapped, mapped in proper pairs, primary alignments, and non-duplicates, (samtools view parameters: -f 2 -F 3972 -q 1), we calculated the coverage with samtools depth. Evenness of coverage was evaluated by calculating the Gini index (80) on the short pDNA and cfDNA's coverage, which was previously binned into adjacent 10 kbp bins.

### ichorCNA

Copy number alterations in the 10X and low-pass WGS were evaluated using ichorCNA (v0.2.0) (39) (github: <https://github.com/broadinstitute/ichorCNA/releases/tag/v0.2.0>).

### Tumor tissue DNA sequencing and analysis

DNA from resected tumor tissue and adjacent tumor-free colonic mucosa was extracted using the Monarch Genomic DNA Purification Kit (Monarch, cat # T3010S). For sequencing, gDNA was prepared as follows: 500 ng per sample at a concentration of 25 ng/µl. The tumor samples were sequenced at 75X and the normal samples at 30X by Novogene UK Ltd. The complete NextFlow pipeline for processing sequencing output to mutation calls is provided in the Zenodo code repository. Briefly, reads were trimmed using Trim Galore! (69) (v0.6.5), requiring >= 2 bp adapter overlap and quality-trimming bases with Phred score below 20, and aligned against the complete hg38 genome, including alt alleles, using bwa-mem2 using default parameters. Mutations were called independently using Mutect2 (v4.1.7.0), Strelka (v2.9.10) and Octopus (v0.7.0). Mutations called by only one caller were excluded.

### Estimation of observed and expected tumor fragments

Ninety-nine sets of random single-base mutations, each matching the unique mutation count identified in the tumor tissue of patient CRC\_012, were generated by sampling from all SNVs detected in any WGS-sequenced colorectal cancer from TCGA. Any SNV overlapping a SNP (dbSNP v155) with > 0.5% allele frequency in 1000Genomes, HapMap, dbGaP\_PopFreq, GnomAD\_exomes, TOPMED or GnomAD were excluded, as well as variants identified as SNPs in the non-cancer control sample for patient CRC\_012. Together with the original

CRC\_012 mutation set, this amounted to 100 mutation sets. For each set, reads covering the mutation positions were extracted, excluding improperly paired reads or reads with mapQ < 25. Detection rate was then calculated by dividing the count of reads containing the variant allele (min base quality: 10) by the total number of reads covering the mutation sites. Positions that exhibited an allele fraction of > 20% in plasma or platelet derived DNA were excluded as potential SNPs. The likelihood of observing a detection rate as high or higher than that of CRC\_012 tissue mutations in that patient's plasma and platelet-derived DNA was calculated using a normal distribution parameterized to the observed mean and standard deviation of detection rates for random mutation sets. Code and intermediate files are deposited in the publication's Zenodo repository.

### TAPS sample preparation

s-pDNA (<600 bp) and l-pDNA (>600 bp) was separated using 0.55X AMPure beads, followed by a 1.8X clean-up of the s-pDNA. Genomic TAPS was carried out on l-pDNA and cfDNA TAPS was carried out on s-pDNA, as previously described (45, 81). Briefly, l-pDNA was mixed with spike-ins of 0.1% CpG-methylated lambda phage DNA and 0.01% unmodified 2 kbp plasmid DNA and was sonicated and size-selected to 300-500 bp prior to library construction. s-pDNA was mixed with pre-fragmented spike-ins of the same ratio and used directly in library construction. DNA was ligated with KAPA Unique Dual-Indexed Adapter Kit using KAPA HyperPrep Kit according to the manufacturer's protocol. The ligated DNA was subjected to mTet1-mediated oxidation and borane reduction. Converted DNA was then amplified with short primers ordered from IDT (FW-5'-AATGATACGGCGACCA CCGAGATCTACAC-3', RV-5'-CAAGCAGAAGACGGCATACGAGAT-3') using KAPA HiFi HotStart Uracil+ ReadyMix PCR Kit for four cycles according to the manufacturer's protocol. The PCR product was purified with AMPure XP beads. Finally, libraries were sequenced for 300 cycles in paired-end mode on the NextSeq 2000 with XLEAP-SBS chemistry P4 cartridge.

### TAPS data processing and analysis

Raw sequencing reads were trimmed from adapter sequences and low-quality bases using Trim Galore! (69) (v0.6.5) and assessed for quality with FastQC (68) (v0.11.9). Reads were aligned to the reference genome using BWA-MEM2 (71) (v2.2.1) and duplicates were marked with Picard (73) (gatk, v4.4.0.0). BAM files were indexed and merged using samtools (72) (v1.16.1). TAPS conversion efficiency and methylation bias were estimated by analyzing sequencing spike-ins. Methylation levels were called using rastair (v0.8.1).

Sample cellular deconvolution was performed using WGBStools and UXM deconv as described in Loyfer *et al.* (2023) (46). A custom cellular atlas was built using the Loyfer *et al.* (2023) pat files for B cells, T cells, NK (natural killer) cells, blood granulocytes, monocytes, erythrocyte progenitors, hepatocytes and vascular endothelium, and an in-house megakaryocyte TAPS dataset. Lymphoid (B cells, T cells, and NK cells) and Myeloid (blood granulocytes, monocytes and erythrocyte progenitors) cells contributions were summed for visualization purposes. All code is available in the Zenodo repository.

### Code availability

The code used in these studies is available from Zenodo (82).

### Statistical analysis

Parametric tests, such as the paired Student's *t* test, were used for comparisons within the same group when the data appeared normally distributed based on visual inspection. When data were paired but not normally distributed, nonparametric tests such as the Wilcoxon signed-rank test were applied. For comparisons between independent groups, the Mann-Whitney *U* test was used to assess statistical significance.

### REFERENCES AND NOTES

- C. Deppermann, P. Kubers, Start a fire, kill the bug: The role of platelets in inflammation and infection. *Innate Immun.* **24**, 335–348 (2018). doi: [10.1177/1753425918789255](https://doi.org/10.1177/1753425918789255); pmid: [30049243](https://pubmed.ncbi.nlm.nih.gov/30049243/)
- J. N. Thon *et al.*, T granules in human platelets function in TLR9 organization and signaling. *J. Cell Biol.* **198**, 561–574 (2012). doi: [10.1083/jcb.201111136](https://doi.org/10.1083/jcb.201111136); pmid: [22908309](https://pubmed.ncbi.nlm.nih.gov/22908309/)
- M. Koupenova *et al.*, The role of platelets in mediating a response to human influenza infection. *Nat. Commun.* **10**, 1780 (2019). doi: [10.1038/s41467-019-09607-x](https://doi.org/10.1038/s41467-019-09607-x); pmid: [30992428](https://pubmed.ncbi.nlm.nih.gov/30992428/)
- M. Koupenova, J. E. Freedman, Platelets and Immunity: Going Viral. *Arterioscler. Thromb. Vasc. Biol.* **40**, 1605–1607 (2020). doi: [10.1161/ATVBAHA.120.314620](https://doi.org/10.1161/ATVBAHA.120.314620); pmid: [32579477](https://pubmed.ncbi.nlm.nih.gov/32579477/)
- M. Koupenova *et al.*, Pollen-derived RNAs Are Found in the Human Circulation. *iScience* **19**, 916–926 (2019). doi: [10.1016/j.isci.2019.08.035](https://doi.org/10.1016/j.isci.2019.08.035); pmid: [31518900](https://pubmed.ncbi.nlm.nih.gov/31518900/)
- R. J. A. Nilsson *et al.*, Blood platelets contain tumor-derived RNA biomarkers. *Blood* **118**, 3680–3683 (2011). doi: [10.1182/blood-2011-03-344408](https://doi.org/10.1182/blood-2011-03-344408); pmid: [21832279](https://pubmed.ncbi.nlm.nih.gov/21832279/)
- Z. Shen *et al.*, Platelet transcriptome identifies progressive markers and potential therapeutic targets in chronic myeloproliferative neoplasms. *Cell Rep. Med.* **2**, 100425 (2021). doi: [10.1016/j.xcrm.2021.100425](https://doi.org/10.1016/j.xcrm.2021.100425); pmid: [34755136](https://pubmed.ncbi.nlm.nih.gov/34755136/)
- M. G. Best *et al.*, RNA-Seq of Tumor-Educated Platelets Enables Blood-Based Pan-Cancer, Multiclass, and Molecular Pathway Cancer Diagnostics. *Cancer Cell* **28**, 666–676 (2015). doi: [10.1016/j.ccell.2015.09.018](https://doi.org/10.1016/j.ccell.2015.09.018); pmid: [26525104](https://pubmed.ncbi.nlm.nih.gov/26525104/)
- S. G. J. G. In 't Veld *et al.*, Detection and localization of early- and late-stage cancers using platelet RNA. *Cancer Cell* **40**, 999–1009.e6 (2022). doi: [10.1016/j.ccell.2022.08.006](https://doi.org/10.1016/j.ccell.2022.08.006); pmid: [36055228](https://pubmed.ncbi.nlm.nih.gov/36055228/)
- C. Bettgeowda *et al.*, Detection of circulating tumor DNA in early- and late-stage human malignancies. *Sci. Transl. Med.* **6**, 224ra24 (2014). doi: [10.1126/scitranslmed.3007094](https://doi.org/10.1126/scitranslmed.3007094); pmid: [24553385](https://pubmed.ncbi.nlm.nih.gov/24553385/)
- A. J. Bronkhorst, V. Ungerer, S. Holdenrieder, The emerging role of cell-free DNA as a molecular marker for cancer management. *Biomol. Detect. Quantif.* **17**, 100087 (2019). doi: [10.1016/j.bdq.2019.100087](https://doi.org/10.1016/j.bdq.2019.100087); pmid: [30923679](https://pubmed.ncbi.nlm.nih.gov/30923679/)
- S. Cristiano *et al.*, Genome-wide cell-free DNA fragmentation in patients with cancer. *Nature* **570**, 385–389 (2019). doi: [10.1038/s41586-019-1272-6](https://doi.org/10.1038/s41586-019-1272-6); pmid: [31142840](https://pubmed.ncbi.nlm.nih.gov/31142840/)
- A. Dasari *et al.*, ctDNA applications and integration in colorectal cancer: An NCI Colon and Rectal-Anal Task Forces whitepaper. *Nat. Rev. Clin. Oncol.* **17**, 757–770 (2020). doi: [10.1038/s41571-020-0392-0](https://doi.org/10.1038/s41571-020-0392-0); pmid: [32632268](https://pubmed.ncbi.nlm.nih.gov/32632268/)
- F. Diehl *et al.*, Circulating mutant DNA to assess tumor dynamics. *Nat. Med.* **14**, 985–990 (2008). doi: [10.1038/nm.1789](https://doi.org/10.1038/nm.1789); pmid: [18670422](https://pubmed.ncbi.nlm.nih.gov/18670422/)
- C. Fiala, E. P. Diamandis, Utility of circulating tumor DNA in cancer diagnostics with emphasis on early detection. *BMC Med.* **16**, 166 (2018). doi: [10.1186/s12916-018-1157-9](https://doi.org/10.1186/s12916-018-1157-9); pmid: [30285732](https://pubmed.ncbi.nlm.nih.gov/30285732/)
- J. C. M. Wan *et al.*, Liquid biopsies come of age: Towards implementation of circulating tumour DNA. *Nat. Rev. Cancer* **17**, 223–238 (2017). doi: [10.1038/nrc.2017.7](https://doi.org/10.1038/nrc.2017.7); pmid: [28233803](https://pubmed.ncbi.nlm.nih.gov/28233803/)
- Y. M. Lo *et al.*, Presence of fetal DNA in maternal plasma and serum. *Lancet* **350**, 485–487 (1997). doi: [10.1016/S0140-6736\(97\)02174-0](https://doi.org/10.1016/S0140-6736(97)02174-0); pmid: [9274585](https://pubmed.ncbi.nlm.nih.gov/9274585/)
- L. Hille *et al.*, Evaluation of an Alternative Staining Method Using SYTO 13 to Determine Reticulated Platelets. *Thromb. Haemost.* **119**, 779–785 (2019). doi: [10.1055/s-0039-1681101](https://doi.org/10.1055/s-0039-1681101); pmid: [30934103](https://pubmed.ncbi.nlm.nih.gov/30934103/)
- P. C. Armstrong *et al.*, Temporal in vivo platelet labeling in mice reveals age-dependent receptor expression and conservation of specific mRNAs. *Blood Adv.* **6**, 6028–6038 (2022). doi: [10.1182/bloodadvances.2022007099](https://doi.org/10.1182/bloodadvances.2022007099); pmid: [36037520](https://pubmed.ncbi.nlm.nih.gov/36037520/)
- H. E. Allan *et al.*, Proteome and functional decline as platelets age in the circulation. *J. Thromb. Haemost.* **19**, 3095–3112 (2021). doi: [10.1111/jth.15496](https://doi.org/10.1111/jth.15496); pmid: [34390534](https://pubmed.ncbi.nlm.nih.gov/34390534/)
- J. Moss *et al.*, Megakaryocyte- and erythroblast-specific cell-free DNA patterns in plasma and platelets reflect thrombopoiesis and erythropoiesis levels. *Nat. Commun.* **14**, 7542 (2023). doi: [10.1038/s41467-023-43310-2](https://doi.org/10.1038/s41467-023-43310-2); pmid: [37985773](https://pubmed.ncbi.nlm.nih.gov/37985773/)
- G. Escolar, E. Leistikow, J. G. White, The fate of the open canalicular system in surface and suspension-activated platelets. *Blood* **74**, 1983–1988 (1989). doi: [10.1182/blood.V74.6.1983.1983](https://doi.org/10.1182/blood.V74.6.1983.1983); pmid: [2804343](https://pubmed.ncbi.nlm.nih.gov/2804343/)
- L. Balaj *et al.*, Tumour microvesicles contain retrotransposon elements and amplified oncogene sequences. *Nat. Commun.* **2**, 180 (2011). doi: [10.1038/ncomms1180](https://doi.org/10.1038/ncomms1180); pmid: [21285958](https://pubmed.ncbi.nlm.nih.gov/21285958/)
- C. Kahler *et al.*, Identification of double-stranded genomic DNA spanning all chromosomes with mutated KRAS and p53 DNA in the serum exosomes of patients with pancreatic cancer. *J. Biol. Chem.* **289**, 3869–3875 (2014). doi: [10.1074/jbc.C113.532267](https://doi.org/10.1074/jbc.C113.532267); pmid: [24398677](https://pubmed.ncbi.nlm.nih.gov/24398677/)
- A. Takahashi *et al.*, Exosomes maintain cellular homeostasis by excreting harmful DNA from cells. *Nat. Commun.* **8**, 15287 (2017). doi: [10.1038/ncomms15287](https://doi.org/10.1038/ncomms15287); pmid: [28508895](https://pubmed.ncbi.nlm.nih.gov/28508895/)
- Z. Hu, H. Chen, Y. Long, P. Li, Y. Gu, The main sources of circulating cell-free DNA: Apoptosis, necrosis and active secretion. *Crit. Rev. Oncol. Hematol.* **157**, 103166 (2021). doi: [10.1016/j.critrevonc.2020.103166](https://doi.org/10.1016/j.critrevonc.2020.103166); pmid: [33254039](https://pubmed.ncbi.nlm.nih.gov/33254039/)
- B. K. Thakur *et al.*, Double-stranded DNA in exosomes: A novel biomarker in cancer detection. *Cell Res.* **24**, 766–769 (2014). doi: [10.1038/cr.2014.44](https://doi.org/10.1038/cr.2014.44); pmid: [24710597](https://pubmed.ncbi.nlm.nih.gov/24710597/)
- D. Jerez-Dolz *et al.*, Internalization of microparticles by platelets is partially mediated by toll-like receptor 4 and enhances platelet thrombogenicity. *Atherosclerosis* **294**, 17–24 (2020). doi: [10.1016/j.atherosclerosis.2019.12.017](https://doi.org/10.1016/j.atherosclerosis.2019.12.017); pmid: [31945614](https://pubmed.ncbi.nlm.nih.gov/31945614/)

29. R. J. Nilsson *et al.*, Rearranged EML4-ALK fusion transcripts sequester in circulating blood platelets and enable blood-based crizotinib response monitoring in non-small-cell lung cancer. *Oncotarget* **7**, 1066–1075 (2016). doi: [10.18632/oncotarget.6279](https://doi.org/10.18632/oncotarget.6279); pmid: [26544515](https://pubmed.ncbi.nlm.nih.gov/26544515/)
30. S. Koseoglu *et al.*, Dynamin-related protein-1 controls fusion pore dynamics during platelet granule exocytosis. *Arterioscler. Thromb. Vasc. Biol.* **33**, 481–488 (2013). doi: [10.1161/ATVBAHA.112.255737](https://doi.org/10.1161/ATVBAHA.112.255737); pmid: [23288151](https://pubmed.ncbi.nlm.nih.gov/23288151/)
31. Y. Yotomi *et al.*, Separable function of platelet release reaction and clot retraction. *Biochem. Biophys. Res. Commun.* **140**, 329–334 (1986). doi: [10.1016/0006-291X\(86\)91094-6](https://doi.org/10.1016/0006-291X(86)91094-6); pmid: [3022719](https://pubmed.ncbi.nlm.nih.gov/3022719/)
32. H. Melchinger, K. Jain, T. Tyagi, J. Hwa, Role of Platelet Mitochondria: Life in a Nucleus-Free Zone. *Front. Cardiovasc. Med.* **6**, 153 (2019). doi: [10.3389/fcvm.2019.00153](https://doi.org/10.3389/fcvm.2019.00153); pmid: [31737646](https://pubmed.ncbi.nlm.nih.gov/31737646/)
33. L. H. Boudreau *et al.*, Platelets release mitochondria serving as substrate for bactericidal group IIA-secreted phospholipase A2 to promote inflammation. *Blood* **124**, 2173–2183 (2014). doi: [10.1182/blood-2014-05-573543](https://doi.org/10.1182/blood-2014-05-573543); pmid: [25082876](https://pubmed.ncbi.nlm.nih.gov/25082876/)
34. S. O'Hagan, M. Wright Muelas, P. J. Day, E. Lundberg, D. B. Kell, GeneGini: Assessment via the Gini Coefficient of Reference "Housekeeping" Genes and Diverse Human Transporter Expression Profiles. *Cell Syst.* **6**, 230–244.e1 (2018). doi: [10.1016/j.cels.2018.01.003](https://doi.org/10.1016/j.cels.2018.01.003); pmid: [29428416](https://pubmed.ncbi.nlm.nih.gov/29428416/)
35. M. W. Snyder, M. Kircher, A. J. Hill, R. M. Daza, J. Shendure, Cell-free DNA Comprises an In Vivo Nucleosome Footprint that Informs Its Tissues-Of-Origin. *Cell* **164**, 57–68 (2016). doi: [10.1016/j.cell.2015.11.050](https://doi.org/10.1016/j.cell.2015.11.050); pmid: [26771485](https://pubmed.ncbi.nlm.nih.gov/26771485/)
36. F. Mouliere *et al.*, Enhanced detection of circulating tumor DNA by fragment size analysis. *Sci. Transl. Med.* **10**, eaat4921 (2018). doi: [10.1126/scitranslmed.aat4921](https://doi.org/10.1126/scitranslmed.aat4921); pmid: [30404863](https://pubmed.ncbi.nlm.nih.gov/30404863/)
37. H. R. Underhill *et al.*, Fragment Length of Circulating Tumor DNA. *PLOS Genet.* **12**, e1006162 (2016). doi: [10.1371/journal.pgen.1006162](https://doi.org/10.1371/journal.pgen.1006162); pmid: [27428049](https://pubmed.ncbi.nlm.nih.gov/27428049/)
38. Y. M. D. Lo, D. S. C. Han, P. Jiang, R. W. K. Chiu, Epigenetics, fragmentomics, and topology of cell-free DNA in liquid biopsies. *Science* **372**, eaaw3616 (2021). doi: [10.1126/science.aaw3616](https://doi.org/10.1126/science.aaw3616); pmid: [33833097](https://pubmed.ncbi.nlm.nih.gov/33833097/)
39. V. A. Adalsteinsson *et al.*, Scalable whole-exome sequencing of cell-free DNA reveals high concordance with metastatic tumors. *Nat. Commun.* **8**, 1324 (2017). doi: [10.1038/s41467-017-00965-y](https://doi.org/10.1038/s41467-017-00965-y); pmid: [29109393](https://pubmed.ncbi.nlm.nih.gov/29109393/)
40. C. Kandoth *et al.*, Mutational landscape and significance across 12 major cancer types. *Nature* **502**, 333–339 (2013). doi: [10.1038/nature12634](https://doi.org/10.1038/nature12634); pmid: [24132290](https://pubmed.ncbi.nlm.nih.gov/24132290/)
41. A. Jamshidi *et al.*, Evaluation of cell-free DNA approaches for multi-cancer early detection. *Cancer Cell* **40**, 1537–1549.e12 (2022). doi: [10.1016/j.ccell.2022.10.022](https://doi.org/10.1016/j.ccell.2022.10.022); pmid: [36400018](https://pubmed.ncbi.nlm.nih.gov/36400018/)
42. M. S. Esfahani *et al.*, Inferring gene expression from cell-free DNA fragmentation profiles. *Nat. Biotechnol.* **40**, 585–597 (2022). doi: [10.1038/s41587-022-01222-4](https://doi.org/10.1038/s41587-022-01222-4); pmid: [35361996](https://pubmed.ncbi.nlm.nih.gov/35361996/)
43. J. Moss *et al.*, Comprehensive human cell-type methylation atlas reveals origins of circulating cell-free DNA in health and disease. *Nat. Commun.* **9**, 5068 (2018). doi: [10.1038/s41467-018-07466-6](https://doi.org/10.1038/s41467-018-07466-6); pmid: [30498206](https://pubmed.ncbi.nlm.nih.gov/30498206/)
44. R. Sadeh *et al.*, ChIP-seq of plasma cell-free nucleosomes identifies gene expression programs of the cells of origin. *Nat. Biotechnol.* **39**, 586–598 (2021). doi: [10.1038/s41587-020-00775-6](https://doi.org/10.1038/s41587-020-00775-6); pmid: [33432199](https://pubmed.ncbi.nlm.nih.gov/33432199/)
45. Y. Liu *et al.*, Bisulfite-free direct detection of 5-methylcytosine and 5-hydroxymethylcytosine at base resolution. *Nat. Biotechnol.* **37**, 424–429 (2019). doi: [10.1038/s41587-019-0041-2](https://doi.org/10.1038/s41587-019-0041-2); pmid: [30804537](https://pubmed.ncbi.nlm.nih.gov/30804537/)
46. N. Loyer *et al.*, A DNA methylation atlas of normal human cell types. *Nature* **613**, 355–364 (2023). doi: [10.1038/s41586-022-05580-6](https://doi.org/10.1038/s41586-022-05580-6); pmid: [36599988](https://pubmed.ncbi.nlm.nih.gov/36599988/)
47. R. Jackstadt *et al.*, Epithelial NOTCH Signaling Rewires the Tumor Microenvironment of Colorectal Cancer to Drive Poor-Prognosis Subtypes and Metastasis. *Cancer Cell* **36**, 319–336.e7 (2019). doi: [10.1016/j.ccell.2019.08.003](https://doi.org/10.1016/j.ccell.2019.08.003); pmid: [31526760](https://pubmed.ncbi.nlm.nih.gov/31526760/)
48. S. Go *et al.*, Tissue-resident NK cells support survival in pancreatic cancer through promotion 1 of cDC1-CD8T activity. *bioRxiv* 2023.10.14.562332 [Preprint] (2024); <https://doi.org/10.1101/2023.10.14.562332>
49. J. E. East *et al.*, British Society of Gastroenterology position statement on serrated polyps in the colon and rectum. *Gut* **66**, 1181–1196 (2017). doi: [10.1136/gutjnl-2017-314005](https://doi.org/10.1136/gutjnl-2017-314005); pmid: [28450390](https://pubmed.ncbi.nlm.nih.gov/28450390/)
50. A. Junca *et al.*, Detection of Colorectal Cancer and Advanced Adenoma by Liquid Biopsy (Decalib Study): The ddPCR Challenge. *Cancers* **12**, 1482 (2020). doi: [10.3390/cancers12061482](https://doi.org/10.3390/cancers12061482); pmid: [32517177](https://pubmed.ncbi.nlm.nih.gov/32517177/)
51. M. Galanopoulos *et al.*, Comparative Study of Mutations in Single Nucleotide Polymorphism Loci of KRAS and BRAF Genes in Patients Who Underwent Screening Colonoscopy, With and Without Premalignant Intestinal Polyps. *Anticancer Res.* **37**, 651–657 (2017). doi: [10.21873/anticancer.11360](https://doi.org/10.21873/anticancer.11360); pmid: [28179313](https://pubmed.ncbi.nlm.nih.gov/28179313/)
52. M. G. Best, P. Wesseling, T. Wurdinger, Tumor-Educated Platelets as a Noninvasive Biomarker Source for Cancer Detection and Progression Monitoring. *Cancer Res.* **78**, 3407–3412 (2018). doi: [10.1158/0008-5472.CAN-18-0887](https://doi.org/10.1158/0008-5472.CAN-18-0887); pmid: [29921699](https://pubmed.ncbi.nlm.nih.gov/29921699/)
53. S. A. Joosse, K. Pantel, Tumor-Educated Platelets as Liquid Biopsy in Cancer Patients. *Cancer Cell* **28**, 552–554 (2015). doi: [10.1016/j.ccell.2015.10.007](https://doi.org/10.1016/j.ccell.2015.10.007); pmid: [26555171](https://pubmed.ncbi.nlm.nih.gov/26555171/)
54. D. C. Calverley *et al.*, Significant downregulation of platelet gene expression in metastatic lung cancer. *Clin. Transl. Sci.* **3**, 227–232 (2010). doi: [10.1111/j.1752-8062.2010.00226.x](https://doi.org/10.1111/j.1752-8062.2010.00226.x); pmid: [21500395](https://pubmed.ncbi.nlm.nih.gov/21500395/)
55. G. H. Frydman *et al.*, Megakaryocytes contain extranuclear histones and may be a source of platelet-associated histones during sepsis. *Sci. Rep.* **10**, 4621 (2020). doi: [10.1038/s41598-020-61309-3](https://doi.org/10.1038/s41598-020-61309-3); pmid: [32165642](https://pubmed.ncbi.nlm.nih.gov/32165642/)
56. L. Clejan, H. Menahem, Binding of deoxyribonucleic acid to the surface of human platelets. *Acta Haematol.* **58**, 84–88 (1977). doi: [10.1159/000207812](https://doi.org/10.1159/000207812); pmid: [409023](https://pubmed.ncbi.nlm.nih.gov/409023/)
57. C. A. Dorsch, Binding of single-strand DNA to human platelets. *Thromb. Res.* **24**, 119–129 (1981). doi: [10.1016/0049-3848\(81\)90037-2](https://doi.org/10.1016/0049-3848(81)90037-2); pmid: [7330833](https://pubmed.ncbi.nlm.nih.gov/7330833/)
58. C. Vanpouille-Box, S. Demaria, S. C. Formenti, L. Galluzzi, Cytosolic DNA Sensing in Organismal Tumor Control. *Cancer Cell* **34**, 361–378 (2018). doi: [10.1016/j.ccell.2018.05.013](https://doi.org/10.1016/j.ccell.2018.05.013); pmid: [30216189](https://pubmed.ncbi.nlm.nih.gov/30216189/)
59. Y. Wang *et al.*, eccDNAs are apoptotic products with high innate immunostimulatory activity. *Nature* **599**, 308–314 (2021). doi: [10.1038/s41586-021-04009-w](https://doi.org/10.1038/s41586-021-04009-w); pmid: [34671165](https://pubmed.ncbi.nlm.nih.gov/34671165/)
60. I. Hawwari *et al.*, Platelet-derived transcription factors licence human monocyte inflammation. *bioRxiv* 2022.08.10.503291 (2023); <https://doi.org/10.1101/2022.08.10.503291>
61. E. S. Kim *et al.*, Blood-based tumor mutational burden as a biomarker for atezolizumab in non-small cell lung cancer: The phase 2 B-FIRST trial. *Nat. Med.* **28**, 939–945 (2022). doi: [10.1038/s41591-022-01754-x](https://doi.org/10.1038/s41591-022-01754-x); pmid: [35422531](https://pubmed.ncbi.nlm.nih.gov/35422531/)
62. D. G. Rothwell *et al.*, Utility of ctDNA to support patient selection for early phase clinical trials: The TARGET study. *Nat. Med.* **25**, 738–743 (2019). doi: [10.1038/s41591-019-0380-z](https://doi.org/10.1038/s41591-019-0380-z); pmid: [31011204](https://pubmed.ncbi.nlm.nih.gov/31011204/)
63. J. D. Merker *et al.*, Circulating Tumor DNA Analysis in Patients With Cancer: American Society of Clinical Oncology and College of American Pathologists Joint Review. *J. Clin. Oncol.* **36**, 1631–1641 (2018). doi: [10.1200/JCO.2017.76.8671](https://doi.org/10.1200/JCO.2017.76.8671); pmid: [29504847](https://pubmed.ncbi.nlm.nih.gov/29504847/)
64. N. Liang *et al.*, Mature Red Blood Cells Contain Long DNA Fragments and Could Acquire DNA from Lung Cancer Tissue. *Adv. Sci. (Weinh.)* **10**, 2206361 (2023). doi: [10.1002/advs.202206361](https://doi.org/10.1002/advs.202206361); pmid: [36599687](https://pubmed.ncbi.nlm.nih.gov/36599687/)
65. C. Schmidt, A. F. Rendeiro, N. C. Sheffield, C. Bock, ChIPmentation: Fast, robust, low-input ChIP-seq for histones and transcription factors. *Nat. Methods* **12**, 963–965 (2015). doi: [10.1038/nmeth.3542](https://doi.org/10.1038/nmeth.3542); pmid: [26280331](https://pubmed.ncbi.nlm.nih.gov/26280331/)
66. J. D. Buenostro, B. Wu, H. Y. Chang, W. J. Greenleaf, ATAC-seq: A Method for Assaying Chromatin Accessibility Genome-Wide. *Curr. Protoc. Mol. Biol.* **109**, 29.1, 9 (2015). doi: [10.1002/0471142727.mb2129s109](https://doi.org/10.1002/0471142727.mb2129s109); pmid: [25559105](https://pubmed.ncbi.nlm.nih.gov/25559105/)
67. Y. Jung, L. Wen, A. Altman, K. Ley, CD45 pre-exclusion from the tips of T cell microvilli prior to antigen recognition. *Nat. Commun.* **12**, 3872 (2021). doi: [10.1038/s41467-021-23792-8](https://doi.org/10.1038/s41467-021-23792-8); pmid: [34162836](https://pubmed.ncbi.nlm.nih.gov/34162836/)
68. S. Andrews, FastQC (Babraham Bioinformatics, Babraham Institute, 2010).
69. F. Krueger, F. James, P. Ewels, E. Aftounian, B. Schuster-Boeckler, FelixKrueger/TrimGalore: v0.6.7, Zenodo (2021); <https://zenodo.org/record/5127899>
70. V. A. Schneider *et al.*, Evaluation of GRCh38 and de novo haploid genome assemblies demonstrates the enduring quality of the reference assembly. *Genome Res.* **27**, 849–864 (2017). doi: [10.1101/gr.213611.116](https://doi.org/10.1101/gr.213611.116); pmid: [28396521](https://pubmed.ncbi.nlm.nih.gov/28396521/)
71. H. Li, Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv:1303.3997 [q-bio.GN]* (2013).
72. H. Li *et al.*, The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**, 2078–2079 (2009). doi: [10.1093/bioinformatics/btp352](https://doi.org/10.1093/bioinformatics/btp352); pmid: [19505943](https://pubmed.ncbi.nlm.nih.gov/19505943/)
73. G. Van der Auwera, B. O'Connor, *Genomics in the cloud: using Docker, GATK, and WDL in Terra* (O'Reilly Media, 2020).
74. H. M. Amemiya, A. Kundaje, A. P. Boyle, The ENCODE Blacklist: Identification of Problematic Regions of the Genome. *Sci. Rep.* **9**, 9354 (2019). doi: [10.1038/s41598-019-45839-z](https://doi.org/10.1038/s41598-019-45839-z); pmid: [31249361](https://pubmed.ncbi.nlm.nih.gov/31249361/)
75. A. R. Quinlan, I. M. Hall, BEDTools: A flexible suite of utilities for comparing genomic features. *Bioinformatics* **26**, 841–842 (2010). doi: [10.1093/bioinformatics/btq033](https://doi.org/10.1093/bioinformatics/btq033); pmid: [20110278](https://pubmed.ncbi.nlm.nih.gov/20110278/)
76. H. Wickham, *ggplot2: Elegant Graphics for Data Analysis* (Springer, 2016), pp. 189–201. doi: [10.1007/978-3-319-24277-4\\_9](https://doi.org/10.1007/978-3-319-24277-4_9)
77. I. Abugessaisa *et al.*, refTSS: A reference data set for human and mouse transcription start sites. *J. Mol. Biol.* **431**, 2407–2422 (2019). doi: [10.1016/j.jmb.2019.04.045](https://doi.org/10.1016/j.jmb.2019.04.045); pmid: [31075273](https://pubmed.ncbi.nlm.nih.gov/31075273/)
78. M. Karimzadeh, C. Ernst, A. Kundaje, M. M. Hoffman, Umap and Bismap: Quantifying genome and methylome mappability. *Nucleic Acids Res.* **46**, e120–e120 (2018). doi: [10.1093/nar/gky677](https://doi.org/10.1093/nar/gky677); pmid: [30169659](https://pubmed.ncbi.nlm.nih.gov/30169659/)
79. B. Gel, E. Serra, karyoploteR: An R/Bioconductor package to plot customizable genomes displaying arbitrary data. *Bioinformatics* **33**, 3088–3090 (2017). doi: [10.1093/bioinformatics/btx346](https://doi.org/10.1093/bioinformatics/btx346); pmid: [28575171](https://pubmed.ncbi.nlm.nih.gov/28575171/)
80. C. Gini, On the measure of concentration with special reference to income and statistics. *Colorado College Publication: General Series* **208**, 73–79 (1936).
81. P. Siejka-Zielińska *et al.*, Cell-free DNA TAPS provides multimodal information for early cancer detection. *Sci. Adv.* **7**, eabh0534 (2021). doi: [10.1126/sciadv.abh0534](https://doi.org/10.1126/sciadv.abh0534); pmid: [34516908](https://pubmed.ncbi.nlm.nih.gov/34516908/)
82. B. Schuster-Boeckler, J. Inchauspe, Scripts and Outputs for Murphy et al. Zenodo (2022).

## ACKNOWLEDGMENTS

We thank J. Koth and C. Lagerholm from the MRC WIMM Imaging Facility; H. Gersen, K. Meissner, F. Roseman, J. Brown, N. Hayder, M. Jacobs, and D. Moralli for ideas and support with experiments; the COMBATcancer consortium; and the CRUK Oxford Centre. We thank the patients and healthy donors for their participation in the research and the Oxford COMBATcancer Consortium and CRUK Oxford Centre for helping with recruitment and sample banking. We also acknowledge BioRender for figure creation. **Funding:** Funding was provided by Cancer Research UK Early Detection Project Grant EDDCPJT/100003 (B.P., C.D.G., P.R.); CRUK Advanced Clinical Scientist Fellowship C67633/A29034 (B.P.); Senior Fellowship RCCSCF-May24/100001 funded by Cancer Research UK in partnership with the Rosetrees Trust (B.P.); Ludwig Institute for Cancer Research Associate Member Program (B.P., B.S.-B.); Medical Research Council Grant MR/T04490X/1 (B.S.-B.); Kidani Memorial Trust (E.O.N.); CRUK Oxford Centre Clinical Research Training Fellowship C130623/A249471 (N.S.); and an unrestricted research grant from Emerson Health (now Yosemite) (B.P.). We also acknowledge support from the National Institute for Health Research (NIHR), Oxford Biomedical Research Centre (BRC), and CRUK Oxford Centre Grant CTRQQR-2021\100002. **Author contributions:** Conceptualization: L.M., B.P., C.D.G.; Methodology: L.M., J.I., N.S., H.L.B.-D., P.H., G.V., R.L., N.J.J., C.B., E.M., Z.E., L.C., E.S., L.D., S.K., I.H., M.K., E.O.N., M.V., P.R., S.L., B.S.-B., C.D.G., B.P.; Software: J.I., B.S.-B., J.R.H.; Validation: L.M., J.I., P.R., C.D.G., B.P., B.S.-B., C.B., N.J.J.; Formal analysis: L.M., J.I., B.S.B., P.R.; Investigation: L.M., J.I., N.S., H.L.B.-D., P.H., G.V., R.L., N.J.J., C.B., S.B., Z.E., L.C., J.O.S., M.P.R., A.O.K., C.S.K., N.N., E.M., E.S., L.D.; Resources: L.M., N.S., B.S.-B., C.D.G., P.R., S.L., M.D., M.V., E.O.N., N.N.; Data curation: L.M., J.I., B.S.-B., J.R.H., Z.E., S.B., L.C., P.R.; Writing - original draft: L.M., B.P.; Writing - review and editing: all authors; Visualization: L.M., J.I., Z.E., L.C., B.S.-B.; Supervision: B.P., A.J.M., P.R., C.D.G., B.S.-B., J.R.H.; Project administration: L.M., B.P., C.D.G., B.S.-B.; Funding acquisition: B.P., C.D.G., B.S.-B. **Competing interests:** B.P., A.J.M., C.D.G., L.M., and N.S. are listed as inventors on patent GB2203947.3 (CRUK Ref: CR/2021-069), submitted in 2022 by the University of Oxford, which covers the detection of disease-associated gene mutations in nucleic acids derived from purified platelets. B.P., A.J.M., and L.M. are also inventors on patent

P45651W01 (GB2314848.9), filed in 2023 by the University of Oxford, relating to the recovery of cell-free nucleic acids from platelets. B.P. is a cofounder and major shareholder of Alethiomics; has received honoraria for speaking engagements and/or advisory work for Incyte, BMS, Constellation Therapeutics, GSK, Blueprint Medicines, Novartis, and Alethiomics; and has received research funding from Incyte, Galecto, and Alethiomics. A.J.M. is a cofounder and shareholder of Alethiomics; has received honoraria or served on advisory boards for Alethiomics, Novartis, CTI, Gilead, and Shire; and has received research funding from Alethiomics and Galecto. L.M. has consulted for Alethiomics. All other authors declare that they have no competing interest. **Data and materials availability:** All data needed to evaluate the conclusions in the paper are present in the paper or the supplementary materials. Raw whole-genome sequencing data has been deposited in the European Genome-phenome Archive (EGA) under accession number EGAS00001006854 (study: EGAS00001006854; dataset: EGAD00001009856; data access committee: EGAC00001003034). Code is available on Zenodo ([82](https://doi.org/10.1126/science.adp3971)). **License information:** Copyright © 2025 the authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works. This work is licensed under a Creative Commons Attribution 4.0 International (CC BY 4.0) license, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. To view a copy of this license, visit <https://creativecommons.org/licenses/by/4.0/>. This license does not apply to figures/photos/artwork or other content included in the article that is credited to a third party; obtain authorization from the rights holder before using such material.

## SUPPLEMENTARY MATERIALS

[science.org/doi/10.1126/science.adp3971](https://doi.org/10.1126/science.adp3971)

Supplementary Text; Figs. S1 to S6; Tables S1 to S7; MDAR Reproducibility Checklist; Movies S1 to S6

Submitted 24 March 2024; resubmitted 1 January 2025; accepted 13 June 2025

10.1126/science.adp3971